1	Title
2	A holocentric twist to chromosomal speciation?
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15	Holocentricity, chromosomal speciation, phylogenetics, karyotype evolution
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18	Abstract
19	Chromosomal rearrangements trigger speciation by acting as barriers to gene flow.
20	However, the underlying theory was developed with monocentric chromosomes in
21	mind. Holocentric chromosomes lacking a centromeric region have repeatedly evolved
22	and account for a significant fraction of extant biodiversity. Because chromosomal
23	rearrangements may be more likely retained in holocentric species, holocentricity could
24	provide a twist to chromosomal speciation. Here we discuss how the abundance of
25	chromosome-scale genomes combined with novel analytical tools offer the opportunity
26	to assess the impacts of chromosomal rearrangements on rates of speciation by
27	outlining a phylogenetic framework that aligns with the two major lines of
28	chromosomal speciation theory. We further highlight how holocentric species could
29	help to test for causal roles of chromosomal rearrangements in speciation.
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A chromosomal view on speciation

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33 While most taxonomic groups have chromosomes with centromeric regions, 34 holocentric (see Glossary) chromosomes that lack such regions have repeatedly 35 evolved in animals and plants [1,2]. Across the tree of life there is moreover a 36 tremendous variation in the number of chromosomes that mono- and holocentric 37 species have, ranging up to three magnitudes of difference within a taxonomic order 38 [3,4]. The evolutionary significance of this variation has gathered much attention over 39 the decades [5–7], and the interest in the evolution of chromosomal changes is currently undergoing a renaissance [8–10]. This is because novel technologies make it possible 40 41 to obtain chromosome-scale genomes even for non-model organisms (e.g. [11,12]). 42 Together with the emergence of new analytical approaches, this allows tackling the 43 evolutionary impact of chromosomal variation, e.g. on rates of speciation [13,14] or 44 gene flow [15]. Variation in chromosome numbers may evolve through very different 45 processes. Large-scale changes in chromosome numbers can for example result from 46 hybridization events [16] or genome duplications through polyploidization, the latter 47 being particularly common in plants [17,18]. Other common processes include the 48 fusion of two chromosomes into a single one or the fission of a chromosome into two, 49 resulting in **dysploidy** [6].

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51 Rearrangements that produce variation in chromosome numbers may eventually 52 result in chromosomal speciation, whereby divergent rearrangements directly or 53 indirectly cause reproductive isolation [5,6]. However, intraspecific karyological 54 variation may also persist and result in only limited levels of reproductive isolation [19-55 22]. Two major lines of theoretical models exist that outline how chromosomal 56 rearrangements could cause chromosomal speciation (reviewed in [7,23]). The first line 57 comprises many of the classic models, which are based on hybrid dysfunction and 58 assume that differentially fixed chromosomal rearrangements between closely related 59 species cause problems during meiosis in hybrids and therefore act as Dobzhansky-60 Muller incompatibilities (DMIs, [5,6,24,25]). The problem with these types of models 61 is that they require chromosomal rearrangements to be fixed in order to be of major 62 effect. This is because newly arising chromosomal rearrangements would typically be 63 underdominant, *i.e.*, they would lead to reduced fitness of hybrid individuals (Fig. 1), 64 either within or between species or populations. While strong underdominance makes

65 it unlikely that novel chromosomal rearrangements spread to fixation, weak 66 underdominance may allow for fixation, but would ensure that chromosomal 67 rearrangements represent only shallow barriers, and are therefore unlikely to cause 68 speciation [7,26]. Nevertheless, empirical evidence for such chromosomal speciation 69 exists, and has been primarily found in mammals [27], including mice [28] and 70 wallabies [8]. Here, monobrachial homology, *i.e.*, multiple chromosomal fusions with 71 one or more common chromosome arms in different fusion arrangements that are fixed 72 between populations or species, has been suggested to result in reproductive isolation 73 [25]. Explanations on how such species may have overcome the underdominance 74 paradox vary, and include genetic drift, genetic bottlenecks and founder effects [29,30]. 75 Indeed, chromosomal speciation may initially result in a reduction of the effective 76 population size (N_e) , which could in turn affect rates of speciation and change the 77 fixation probabilities of new karyotypes in allopatry [31]. This has been suggested for 78 mammals, where families with large geographic distributions but whose species have 79 restricted geographic ranges showed a greater probability for fixing different 80 karyotypes [32]. Shifts in mating system, e.g., from outcrossing to selfing [33] or 81 meiotic drive, whereby some alleles or associated rearrangements are more likely to 82 be transmitted [34] have similarly been suggested to overcome underdominance. All 83 these scenarios have received much criticism in the past, however, and formal 84 experiments for a causal association between chromosomal rearrangements and 85 speciation are lacking [7,24].

86 The second major line of theoretical models was developed more recently and 87 has attempted to overcome the underdominance paradox by focusing on changes in 88 recombination associated with chromosomal rearrangements [7,23,26,35]. In essence, 89 under these suppression of recombination type models, rearranged chromosomes can 90 become fixed by drift but also by selection, e.g., when two or more adaptive loci 91 become physically coupled or by locally reducing recombination, both enhancing 92 existing reproductive isolation [26,35]. Such rearranged regions of reduced 93 recombination may act as barrier loci and promote further differentiation, which may 94 eventually lead to postzygotic isolation through the buildup of genetic incompatibilities 95 [7,26,35]. Reproductive isolation associated with chromosomal rearrangements may be 96 further enhanced by sexual selection or reinforcement and may thus promote 97 speciation upon secondary contact. If chromosomal rearrangements contain physically 98 linked clusters of genes they may themselves represent genomic islands of 99 differentiation, or **supergenes** [36,37]. Albeit such supergenes have been suggested to 100 promote speciation [38], their actual contribution towards reproductive isolation 101 remains controversial [39].

102 The current theory on chromosomal speciation has an important gap – it is based 103 on the assumption that chromosomes are monocentric and have a centromeric region 104 that concentrates all kinetochores for the attachment of the spindle tubules during 105 mitosis and meiosis [5-7] (Fig. 2). However, holocentric chromosomes that lack a 106 centromeric region have evolved in very distinct taxonomic groups (Fig. 3), comprising 107 some of the most diverse branches of the tree of life such as the sedge family 108 Cyperaceae with ~5'500 species [40], the order Lepidoptera with ~160'000 butterfly 109 and moth species [41], as well as the nematode model organism *Caenorhabditis elegans* 110 [2]. In contrast to monocentric chromosomes, holocentric chromosomes have 111 molecular features that allow kinetochore proteins to bind along the entire chromosome, 112 permitting microtubules to attach broadly [1] (Fig. 2). As a consequence, rearranged 113 parts of the genome may not cause segregation problems during cell divisions. 114 Holocentricity could therefore provide a twist to chromosomal speciation theory. 115 Indeed White already highlighted in his classic work on chromosomal speciation [5] 116 that "The laws and principles of chromosomal rearrangements in these [holocentric] 117 organisms are not yet fully understood, but certainly they differ in some respects from 118 those governing chromosomal rearrangements in species with the more usual 119 monocentric chromosome." However, despite a recent increase in interest in the 120 evolutionary implications of holocentric chromosomes, the potential effects of 121 holocentricity on chromosomal speciation have remained unclear [2,9,10]. 122 Holocentricity may for example help to overcome the initial underdominance paradox 123 of the classic chromosomal speciation theory (Fig 1). This is because large-scale 124 rearrangements through chromosomal fusions as well as fissions may be more likely to 125 be retained as rearranged chromosomes maintain kinetochore function [1]. This 126 contrasts to most scenarios in monocentric species, where fission events result in 127 chromosomal segments that are not attached to a centromere and may therefore be lost 128 during meiosis (Fig. 2) or where fusion events result in dicentric chromosomes with 129 two centromeres and similarly cause problems during meiosis [42]. Monocentric 130 chromosomal fusions may not always result in segregation problems though, *e.g.* when

131 two chromosomes with terminal centromeres are involved and both chromosomal arms 132 are retain in the fused chromosome [25]. This scenario applies, however, only when 133 nearly complete chromosomes become rearranged and excludes fission events. In 134 addition, intraspecific crosses between holocentric chromosomal races or closely 135 related species may not necessarily cause a significant immediate reduction in offspring 136 fitness [19-22], suggesting that suppression of recombination could also be an 137 important driver of chromosome associated speciation in taxa with holocentric 138 chromosomes.

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140 Mono- and holocentric species further differ in several aspects of their meiotic 141 cell division that may affect the potential for chromosomal speciation. In holocentric 142 species the recombination and segregation functions interfere during meiosis, 143 restricting the potential number of chiasmata in bivalents [43]. In this way, some 144 holocentric groups have evolved an inverted meiosis, where, opposite to monocentric 145 groups, the first meiotic division separates the sister chromatids and the second division 146 the chromosomal homologs [19,21]. This inverted meiosis has been suggested to 147 promote the evolution of new karyotypes and possibly chromosomal speciation by 148 facilitating a correct chromosome segregation in hybrids between populations or 149 species that differ in their karyotype [21]. Other holocentric groups, like the nematode 150 C. elegans, have evolved a monokinetic-like meiosis as they only keep kinetochore 151 activity in the telomeres [44] avoiding potential interference between chiasmata and 152 spindles. While these mechanisms may help to establish novel karyotypes, their impact 153 on meiotic recombination remains unclear [45], also because comparatively few 154 recombination maps exist so far for holocentric species [45-47]. Importantly, because 155 recombination is often, but not always [47], reduced close to centromeres in 156 monocentrics [46], patterns of recombination are likely to differ across holocentric 157 chromosomes. Also, while holocentric chromosomes lack a centromere, their 158 kinetochores may not be equally distributed [48]. The latter is true for C. elegans [44], 159 where recombination increases towards the telomeric regions in contrast to the postman 160 butterfly Heliconius melpomene, where recombination is similar across chromosomes 161 [47]. Processes similar to meiotic drive in monocentric species may consequently be at 162 play for holocentric species as has been found for sedges, rushes (Juncus sp.) and other 163 holocentric lineages [49]. However, given the repeated evolution of holocentricity, it remains to show to which degree such holokinetic drive may be common amongholocentric groups.

166

167 The causality between chromosomal rearrangements and species diversification 168 has remained contentious [7]. Phylogenetic inferences suggest that rates of 169 chromosome evolution might be similar between holo- and monocentric species in 170 insects [9], and that rates of diversification are similar between holo- and monocentric 171 clades when comparing sister holo- and monocentric taxonomic orders across 172 eukaryotes [50]. However, there is often substantial variation in chromosome numbers 173 between genera or families within orders, that are moreover associated with different 174 rates of speciation [8,10,51]. The relative contribution of chromosomal fusion and 175 fission on phylogenetic species diversification varies similarly among taxonomic 176 groups and thus likely impacts rates of diversification differently [10,51]. Empirical 177 evidence for chromosomal speciation is rare, either because speciation is already 178 complete or not, often precluding causal implications of one or multiple rearrangements 179 [7,19]. The few examples for holocentric species suggest that intrinsic postzygotic 180 reproductive isolation between species with different karyotype seems to be limited for 181 Lepidoptera [16,19,22,52] and sedges [53,54]. Experimental hybrids between 182 cytogenetic races of the same sedge species showed that hybrid dysfunction is very 183 limited between populations that differ in few chromosome rearrangements but 184 increases as the number of chromosome rearrangements increase [53,54]. These few 185 empirical examples contrast the vast diversity of the taxonomic groups that have 186 evolved holocentric chromosomes and karyotypic diversity [1,2,9]. Here, novel 187 phylogenetic approaches [13,14] could help to assess the macroevolutionary 188 implications of changes in chromosome numbers more generally and provide a 189 framework for comparative analyses between holo- and monocentric groups.

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191 A phylogenetic framework of chromosomal speciation

Recent advances allow to disentangle models of chromosomal evolution in a phylogenetic framework and to distinguish if a phylogenetic event is rather associated with **ana**- or **cladogenesis** [13,14] (Fig. 4). Under cladogenesis, karyotype evolution occurs at a speciation event, while under anagenesis karyotypes evolve along a branch and speciation happens later. Ana- and cladogenesis are compatible with the two aforementioned major lines of chromosomal speciation models, where cladogenesis resembles the classic hybrid dysfunction type models and anagenesis the recombination
suppression type models. Importantly, ana- and cladogenetic processes may not be
exclusive and may similarly result in a phylogenetic event when they occur together.

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202 The phylogenetic framework outlined in (Fig. 4) allows to quantify how 203 common changes in karyotype numbers might be associated with speciation events at 204 a macroevolutionary scale and to compare between mono- and holocentric clades [9]. 205 Current limitations are primarily given by the availability of dense phylogenies 206 associated with associated karyotype data, often only allowing to study chromosomal 207 speciation at a lower taxonomic level [10]. As this framework allows identifying 208 branching events that are more likely to have resulted in ana- or cladogenetic events 209 respectively, such species pairs could be used to perform in-depth comparative genomic 210 analyses to identify which rearrangements are more likely to result in one or the other 211 phylogenetic event.

212

213 Because a correct segregation of chromosomes may initially be often possible 214 in hybrids of holocentric parental species with different karyotypes [21], holocentricity 215 provides an excellent system to experimentally study chromosomal speciation. The 216 outlined phylogenetic analyses (Fig. 4) combined with crossing experiments could for 217 example quantify the impact of chromosomal rearrangements on reproductive isolation 218 in relation to *e.g.*, the respective evolutionary distance among distinct species pairs. 219 Irradiation based experiments on holocentric plants moreover suggest that 220 holocentricity and a fast formation of new telomeres at breakpoints enables rapid 221 karyotype evolution in holocentric species, though the impact on reproductive isolation 222 was not tested [55]. As direct experimental manipulations of individual chromosomes 223 become technically feasible through novel laser nanosurgery approaches [56] or by 224 generating artificial chromosomes [57], the outcome of specific artificial fusion or 225 fission events can now be experimentally studied, enabling to recreate karyotypic 226 changes between sibling species and to assess their direct impact on reproductive 227 isolation.

228

229 Concluding remarks

Although some of the most diverse taxonomic groups of animals and plants haveevolved holocentric chromosomes [1], the potential evolutionary implications of

232 holocentricity remain elusive. As we outlined, holocentricity could provide a new twist 233 to chromosomal speciation but further research is required. Studying holocentric 234 species could help to advance our understanding on chromosomal speciation (see 235 Outstanding Questions). In addition, we suggest future theoretical explorations, for 236 example, to assess the potential for chromosomal speciation in holocentric taxa, where 237 novel rearrangements may not immediately result in hybrid dysfunction but include a 238 lag time during which heterozygous rearrangements may be tolerated [19]. While 239 chromosomal rearrangements could be an important driver of speciation in cases where 240 they contribute to reproductive isolation, additional pre-zygotic barriers may need to 241 subsequently evolve to complete the speciation process [7]. Comparisons between 242 evolutionary young sibling species that coexist or form zones of secondary contact are 243 thus needed to assess the contribution of chromosomal rearrangements on reproductive 244 isolation in relation to other barriers [58]. To further gain a better understanding on the 245 macroevolutionary impact of karyotype evolution and holocentricity more in-depth and 246 comparative analyses are required to first identify the genomic mechanisms underlying 247 chromosomal fusion and fission sites and their (non-)parallelism across holocentric 248 groups. This would then allow us to identify why for example in Lepidoptera only some 249 genera show tremendous karyotypic variation whereas other genera show none [10]. 250 Lastly, the increased availability of genomic resources for non-model species combined 251 with recently developed models for chromosome evolution [13,14] allow for large-252 scale macroevolutionary studies both within and across taxonomic orders to decipher 253 the evolutionary consequences of holocentricity.

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266 **References**

- Melters, D.P. *et al.* (2012) Holocentric chromosomes: convergent evolution, meiotic adaptations, and genomic analysis. *Chromosome Res.* 20, 579–593.
- 269 2. Mandrioli, M. and Manicardi, G.C. (2020) Holocentric chromosomes. *PLOS Genet.* 16, e1008918. DOI: 10.1371/journal.pgen.1008918
- 271 (https://journals.plos.org/plosgenetics)
- 272 3. Rice, A *et al.* (2015) The chromosome counts database CCDB a community resource of plant chromosome numbers. *New Phytol.* 206, 19–26.
- 4. Román- Palacios, C. *et al.* (2021) Animal chromosome counts reveal a similar range of chromosome numbers but with less polyploidy in animals compared to flowering plants. *J. Evol. Biol.* 34, 1333–1339.
- 5. White, M. J. D. (1978) *Modes of Speciation*, W. H. Freeman and Company.
- King, M. (1995) Species Evolution: The Role of Chromosome Change, Cambridge
 University Press.
- Faria, R. and Navarro, A. (2010) Chromosomal speciation revisited: rearranging
 theory with pieces of evidence. *Trends Ecol. Evol.* 25, 660–669.
- Potter, S. *et al.* (2017) Chromosomal speciation in the genomics era:
 disentangling phylogenetic evolution of rock-wallabies. *Front. Genet.* DOI:
 10.3389/fgene.2017.00010 (https://www.frontiersin.org/journals/genetics)
- 9. Ruckman, S.N. *et al.* (2020) Chromosome number evolves at equal rates in holocentric and monocentric clades. *PLOS Genet*. DOI:
- 287 10.1371/journal.pgen.1009076 (https://journals.plos.org/plosgenetics)
- 288 10. de Vos, J.M. *et al.* (2020) Speciation through chromosomal fusion and fission in
 289 Lepidoptera. *Phil. Trans. R. Soc. B* 375, 20190539.
- 290 11. Cicconardi, F. *et al.* (2021) Chromosome fusion affects genetic diversity and
 291 evolutionary turnover of functional loci, but consistently depends on chromosome
 292 size. *Mol. Biol. Evol.* 10, 4449–4462.
- 293 12. Rhie, A. *et al.* (2021) Towards complete and error-free genome assemblies of all vertebrate species. *Nature* 592, 737–746.
- Freyman, W.A. and Höhna, S. (2018) Cladogenetic and anagenetic models of
 chromosome number evolution: a Bayesian model averaging approach. *Syst. Biol.*67, 195–215.
- 14. Mayrose, I. and Lysak, M.A. (2021) The evolution of chromosome numbers:
 mechanistic models and experimental approaches. *Genome Biol. Evol.* DOI
 10.1093/gbe/evaa220 (https://academic.oup.com/gbe)
- 301 15. Patterson, N. *et al.* (2012) Ancient admixture in human history. *Genetics* 192, 1065–1093.
- 16. Lukhtanov, V.A. *et al.* (2015) Homoploid hybrid speciation and genome
 evolution via chromosome sorting. *Proc. R. Soc. B.* 282, 20150157.
- 305 17. Mayrose, I. *et al.* (2010) Probabilistic models of chromosome number evolution
 306 and the inference of polyploidy. *Syst. Biol.* 59, 132–144.
- 307 18. Rieseberg, L.H. and Willis, J.H. (2007) Plant speciation. Science 317, 910–914.
- 308 19. Lukhtanov, V.A. *et al.* (2020) Incomplete sterility of chromosomal hybrids:
 309 implications for karyotype evolution and homoploid hybrid speciation. *Front.*310 *Genet.* DOI:10.3389/fgene.2020.583827
- 311 (https://www.frontiersin.org/journals/genetics)
- 20. Escudero, M. *et al.* (2013) Species coherence in the face of karyotype
 diversification in holocentric organisms: the case of a cytogenetically variable
 sedge (*Carex scoparia*, Cyperaceae). *Ann. Bot.* 112, 515–526.

315 21. Lukhtanov, V.A. et al. (2018) Versatility of multivalent orientation, inverted 316 meiosis, and rescued fitness in holocentric chromosomal hybrids. Proc. Natl. 317 Acad. Sci. U.S.A. 115, E9610-E9619. 318 22. Hora, K.H. et al. (2019) Limited intrinsic postzygotic reproductive isolation despite chromosomal rearrangements between closely related sympatric species of 319 320 small ermine moths (Lepidoptera: Yponomeutidae). Biol. J. Linn. Soc. 128, 44-321 58. 322 23. Rieseberg, L.H. (2001) Chromosomal rearrangements and speciation. Trends 323 Ecol. Evol. 16, 351-358. 324 24. Coyne, J.A. and Orr, H.A., eds (2004) Speciation, Oxford University Press. 325 25. Baker, R.J. and Bickham, J.W. (1986) Speciation by monobrachial centric 326 fusions. Proc. Natl. Acad. Sci. U.S.A. 83, 8245-8248. 327 26. Navarro, A. and Barton, N.H. (2003) Accumulating postzygotic isolating genes in 328 parapatry: a new twist on chromosomal speciation. Evolution 57, 447-459. 329 27. Bush, G.L. et al. (1977) Rapid speciation and chromosomal evolution in 330 mammals. Proc. Natl. Acad. Sci. U.S.A. 74, 3942-3946. 331 28. Garagna, S. et al. (2014) The Robertsonian phenomenon in the house mouse: 332 mutation, meiosis and speciation. Chromosoma 123, 529-544. 333 29. Lande, R. (1979) Effective deme sizes during long-term evolution estimated from 334 rates of chromosomal rearrangement. Evolution 33, 234–251. 335 30. Templeton, A.R. (1981) Mechanisms of speciation - a population genetic 336 approach. Annu. Rev. Ecol. Evol. Syst. 12, 23-48. 337 31. Lanfear, R. (2014) Population size and the rate of evolution. Trends Ecol. Evol. 338 29, 33-41. 339 32. Martinez, P.A. et al. (2017) A comparative study on karyotypic diversification 340 rate in mammals. Heredity 118, 366-373. 341 33. Robertson, K. et al. (2011) Comparative evidence for the correlated evolution of 342 polyploidy and self-compatibility in Solanaceae. Evolution 65, 139–155. 343 34. de Villena, F.P. and Sapienza, C. (2001) Female meiosis drives karyotypic 344 evolution in mammals. Genetics 159, 1179-1189. 345 35. Guerrero, R.F. and Kirkpatrick, M. (2014) Local adaptation and the evolution of 346 chromosome fusions. Evolution 68, 2747-2756. 347 36. Wellenreuther, M. and Bernatchez, L. (2018) Eco-evolutionary genomics of 348 chromosomal inversions. Trends Ecol. Evol. 33, 427-440. 349 37. Yeaman, S. (2013) Genomic rearrangements and the evolution of clusters of 350 locally adaptive loci. Proc. Natl. Acad. Sci. U.S.A. 110, E1743-E1751. 351 38. Feder, J.L. et al. (2012) The genomics of speciation-with-gene-flow. Trends 352 Genet. 28, 342-350. 353 39. Ravinet, M. et al. (2017) Interpreting the genomic landscape of speciation: a road 354 map for finding barriers to gene flow. J. Evol. Biol. 30, 1450–1477. 355 40. Larridon, I. et al. (2021) A new classification of Cyperaceae (Poales) supported by phylogenomic data. J. Syst. Evol. 59, 852-895. 356 357 41. Stork, N.E (2018) How many species of insects and other terrestrial arthropods 358 are there on Earth? Annu. Rev. Entomol. 63, 31-45. 359 42. Stimpson, K.M. et al. (2012) Dicentric chromosomes: unique models to study 360 centromere function and inactivation. Chromosome Res. 20, 595-605. 361 43. Nokkala, S. et al. (2004) Holocentric chromosomes in meiosis. I. Restriction of 362 the number of chiasmata in bivalents. Chromosome Res, 12, 733-739. 363 44. Maddox, P.S. et al. (2004) 'Holo'er than thou: chromosome segregation and 364 kinetochore function in C. elegans. Chromosome Res. 12, 641–653.

365	45.	Hofstatter, P.G. et al. (2021) Meiosis progression and recombination in
366		holocentric plants: what is known? Front. Plant Sci. DOI:
367		10.3389/fpls.2021.658296 (https://www.frontiersin.org/journals/plant-science)
368	46.	Stapley, J. et al. (2017) Variation in recombination frequency and distribution
369		across eukaryotes: patterns and processes. Phil. Trans. R. Soc. B 372, 20160455.
370	47.	Haenel, Q. et al. (2018) Meta-analysis of chromosome-scale crossover rate
371		variation in eukaryotes and its significance to evolutionary genomics. <i>Mol. Ecol.</i>
372		27, 2477–2497.
373	48.	Senaratne, A.P. et al. (2021) Formation of the CenH3-deficient holocentromere in
374		Lepidoptera avoids active chromatin. Curr. Biol. 31, 173-181.
375	49.	Krátká, M. et al. (2021) Holocentric chromosomes probably do not prevent
376		centromere drive in Cyperaceae. Front. Plant Sci. DOI: 10.3389/fpls.2021.642661
377		(https://www.frontiersin.org/journals/plant-science)
378	50.	Márquez-Corro, J.I. <i>et al.</i> (2018) Do holocentric chromosomes represent an
379		evolutionary advantage? A study of paired analyses of diversification rates of
380		lineages with holocentric chromosomes and their monocentric closest relatives.
381		Chromosome Res. 26, 139–152.
382	51	Márquez- Corro, J.I. <i>et al.</i> (2021) Macroevolutionary insights into sedges (<i>Carex</i> :
383	01.	Cyperaceae): The effects of rapid chromosome number evolution on lineage
384		diversification. J. Syst. Evol. 59, 776–790.
385	52	Wadsworth, C.B. <i>et al.</i> (2015) A recombination suppressor contributes to
386	52.	ecological speciation in OSTRINIA moths. Heredity 114, 593–600.
387	53	Escudero, M. <i>et al.</i> (2016) Chromosomal rearrangements in holocentric organisms
388	55.	lead to reproductive isolation by hybrid dysfunction: The correlation between
389		karyotype rearrangements and germination rates in sedges. Am. J. Bot. 103, 1529–
390		1536.
391	54	Whitkus, R. (1988) Experimental hybridizations among chromosome races of
392	54.	<i>Carex pachystachya</i> and the related species <i>C. macloviana</i> and <i>C. preslii</i>
393		(Cyperaceae). Syst. Bot. 13, 146-153.
394	55	Jankowska, M. <i>et al.</i> (2015) Holokinetic centromeres and efficient telomere
395	55.	healing enable rapid karyotype evolution. <i>Chromosoma</i> 124, 519–528.
396	56	Blázquez-Castro, A. <i>et al.</i> (2020) Genetic material manipulation and modification
397	50.	by optical trapping and nanosurgery - a perspective. <i>Front. Bioeng. Biotechnol.</i>
398		DOI: 10.3389/fbioe.2020.580937
399		
400	57	(https://www.frontiersin.org/journals/bioengineering-and-biotechnology) Lin, Z. <i>et al.</i> (2021) Formation of artificial chromosomes in <i>Caenorhabditis</i>
400	57.	<i>elegans</i> and analyses of their segregation in mitosis, DNA sequence composition
401		
	50	and holocentromere organization. <i>Nucleic Acids Res.</i> 49, 9174–9193.
403	38.	Kulmuni, J. <i>et al.</i> (2020) Towards the completion of speciation: the evolution of
404		reproductive isolation beyond the first barriers. <i>Phil. Trans. R. Soc. B</i> 375, 20100528
405	50	20190528. Lukhtaran V (2015) The blue butterfly Behavior star (Blahieule) etlevitions
406	39.	Lukhtanov, V. (2015) The blue butterfly <i>Polyommatus (Plebicula) atlanticus</i>
407		(Lepidoptera, Lycaenidae) holds the record of the highest number of
408		chromosomes in the non-polyploid eukaryotic organisms. <i>Comp Cytogenet</i> . 9,
409	()	683–690.
410	60.	Koren, T. and Lauš, B. (2015) The Grecian anomalous blue <i>Polyommatus</i>
411		(Agrodiaetus) aroaniensis (Brown, 1976) (Lepidoptera: Lycaneidae) discovered
412		in Croatia, at the north-western edge of its distribution. Nat. Slov. 17, 47-57.

- 61. Vishnevskaya, M.S. *et al.* (2016) Karyosystematics and molecular taxonomy of the anomalous blue butterflies (Lepidoptera, Lycaenidae) from the Balkan
- Peninsula. Comp Cytogenet. 10, 1–85.
- 417

418 Figure legends

Fig. 1: The underdominance paradox. Depicted are hypothetical fitness landscapes for species with different karyotypes and their F1 hybrids with the effects of strong and weak underdominance as predicted by theory for monocentric species [7,26]. While no such theory exists for holocentric species, predictions are given based on empirical findings, which suggest that F1 hybrids in holocentric species may not necessarily suffer from the underdominance paradox [19,21].

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Fig. 2: Comparison of the outcomes of fission events during cell division for mono- and holocentric species. If fission occurs during anaphase, the fragment that is not attached to a centromere is lost for monocentric species. In contrast, fragmented chromosome sections of holocentric species can maintain kinetochore function due to the distribution of centromere-like structures along the chromosome, and may so be retained.

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Fig. 3: Examples of the diversity of holocentric species and their haploid karyotypes. A – *Carex esenbeckiana* (n = 13). B – *C. fischeri* (n = 36). C – *Polyommatus atlantica* (n = 224), adapted from the Natural History Museum London & [59]. D – *P. aroaniensis* (n = 47), adapted from [60], [61]. Pictures in A, B – courtesy of Modesto Luceño Garces. Scale bars represent a length of 10 μ m.

437

Fig. 4: Contrasting phylogenetic models of karyotype evolution with their putative counterparts of major lines of chromosomal speciation models. The outcome of the different models of karyotype evolution are outlined along a hypothetical phylogeny, with clado- and/or anagenetic karyotypical changes being indicated. Colors of branches indicate changes in haploid chromosome numbers, while color gradients indicate that the process of karyotype fixation may occur more slowly after anagenetic changes.

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- 451 Glossary
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453 Anagenesis: Type of speciation in which an ancestral species gradually evolves
454 into another by accumulating changes within a single lineage over time.

455 Artificial chromosomes: Artificially created chromosomes that have the 456 necessary properties (e.g. centromeres, telomeres and origins of replication) to be self-457 replicating and stable.

458 Dobzhansky-Muller incompatibilities: Negative epistatic interactions or
459 incompatibilities that occur between loci with different evolutionary histories.
460 Populations may diverge in allopatry and accumulate such incompatibilities through
461 drift and/or through mutations that prevent hybridization upon secondary contact.

462 **Dysploidy**: Process that increases or decreases the number of chromosomes
463 within a species through chromosomal rearrangements with no significant changes in
464 DNA content.

465 Chiasma: Point of contact between chromatids from two homologous
466 chromosomes during meiotic divisions that allows recombination through
467 chromosomal crossovers between both chromatids.

468 Cladogenesis: Type of speciation in which an ancestral species splits into two469 or more species.

470 Genetic drift: A stochastic evolutionary process that results in changes of allele471 frequencies by sampling a finite number of individuals each generation.

472 Holocentric / holokinetic chromosome: Chromosomes with non-localized
473 centromere-like structures. The kinetochore activity is distributed along the whole
474 chromosome.

475 Holokinetic drive: Perturbation of the normal meiotic process so that a
476 particular allele is preferentially transmitted to the progeny over another allele caused
477 by variation in kinetochore distribution along the holocentric chromosomes or the size
478 of holocentric chromosomes.

479 Kinetochores: Protein structures located on the chromosomes. Microtubules of480 the mito- or meiotic spindles are anchored to this structure during cell division. For

481 monocentric species, kinetochores are located in the centromere whereas for482 holocentric species, they occur throughout the chromosomes.

483 Meiotic drive: Perturbation of the normal meiotic process so that a particular
484 allele is preferentially transmitted to the progeny over another allele. The centromere,
485 its location and size are factors that can result in meiotic drive.

486 Monobrachial homology: Homology between two bi-armed chromosomes that487 is restricted to only one of the two chromosome arms.

488 Polyploidy: Chromosome multiplication entailing the addition of complete489 chromosome sets.

490 Reinforcement: Evolutionary process whereby pre- or postzygotic mechanisms
 491 increase reproductive isolation between two closely related lineages upon secondary
 492 contact.

493 Supergene: A set of genes in strong linkage that segregate together during
494 meiotic divisions because there is a mechanism that impedes recombination within the
495 supergene, such as chromosomal rearrangements, like inversions.

496 Underdominance: Strong selection against heterozygotes. For chromosomally
497 diverging populations, chromosomal hybrids have low fitness and there is a strong
498 selection against them.

499

500

Highlights

Chromosomal speciation, whereby major chromosomal rearrangements trigger reproductive isolation, is a classic evolutionary concept.

The underlying theory was developed for chromosomes with centromeres when holocentric chromosomes that lack centromeres have repeatedly evolved across the tree of life.

We argue that holocentricity may help to overcome problems associated with classic chromosomal speciation theory and that the special characteristics of holocentric chromosomes vastly expand the potential for experimental research on chromosomal speciation.

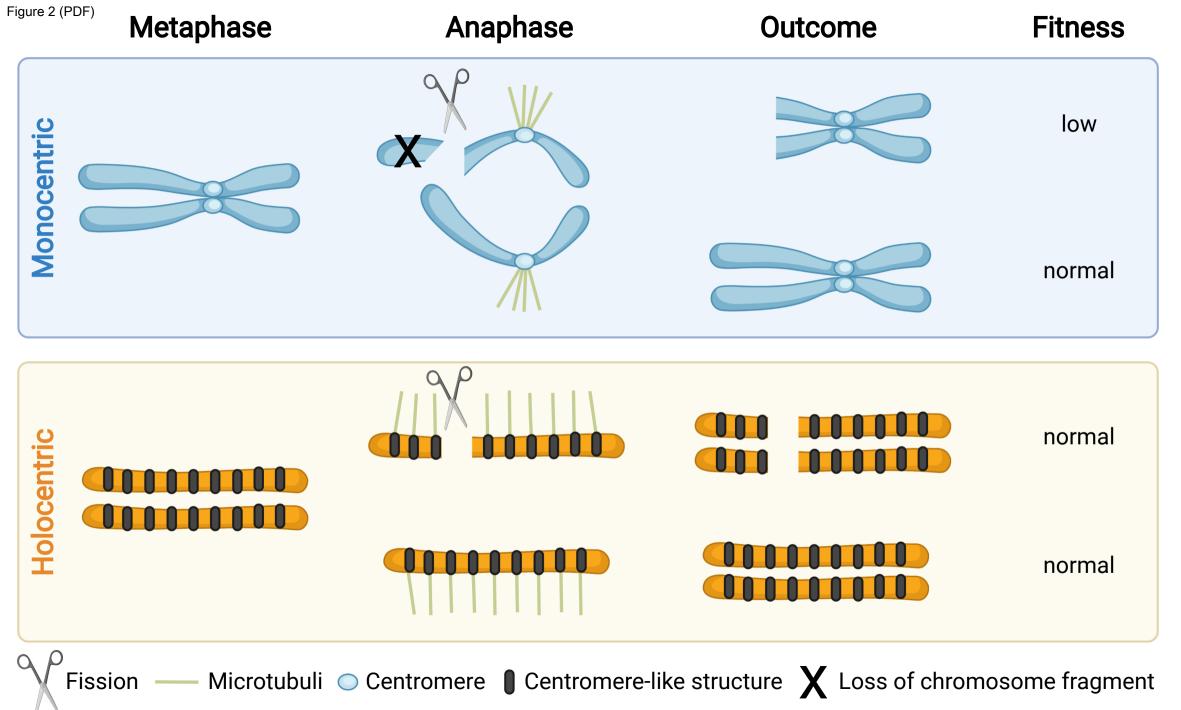
We outline how new approaches allow to quantify the macroevolutionary impact of chromosomal speciation and to distinguish the associated evolutionary mechanisms.

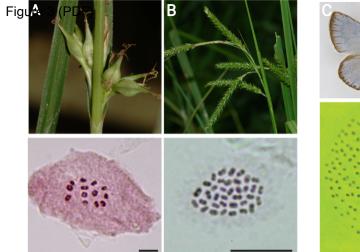
Outstanding questions

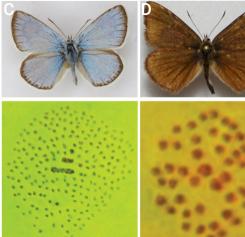
- What are the genomic features underlying chromosomal fusion and fission sites and did they evolve repeatedly across the tree of life? Are there common rearrangement hotspots?
- How do chromosomal rearrangements affect gene flow and does it differ between mono- and holocentric species?
- If rearranged chromosomes act as barrier loci, how does reproductive isolation buildup in the rest of the genome? Are rearranged regions enriched for functional genes?
- How does recombination differ between mono- and holocentric species and what are the implications of fusion and fission on recombination?
- To which degree do ana- and cladogenic phylogenetic events reflect the two lines of chromosomal speciation theory?
- What is the macroevolutionary impact of chromosomal rearrangements between mono- and holocentric species and what are the predominant underlying mechanisms? (see Box 1)
- Is chromosomal speciation more likely to occur in holocentric species?

Figure 1 (PDF)

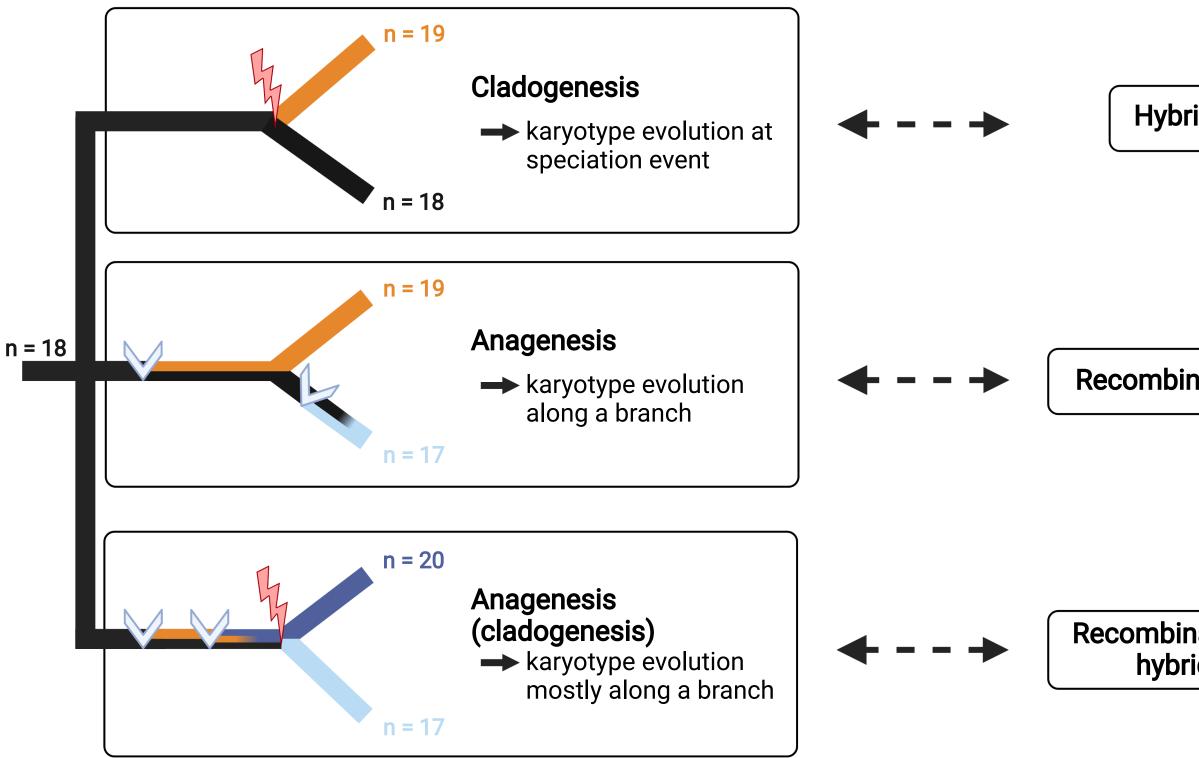
igure i (FDF)	Eitness F1	Fitness	F1
	Monocentric	Monocentric	Holocentric
Underdominance	strong	weak	weak
Establishment of new karyotype	hard	easy	easy
Reproductive barrier strength	strong	weak	weak
F1 fitness	low	normal	normal







Phylogenetic models of trait evolution





Anagenetic chromosomal change

n = Haploid chromosome number

Chromosomal speciation models

Hybrid dysfunction

Recombination suppression

Recombination suppression/ hybrid dysfunction