Mono-alkyl Phosphinic Acids as Ligands in Nanocrystal Synthesis

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Abstract

Ligands play a crucial role in the synthesis of colloidal nanocrystals. Nevertheless, only a handful molecules are currently used, oleic acid being the most typical example. Here, we show that mono-alkyl phosphinic acids are another interesting ligand class, forming metal complexes with a reactivity that is intermediate between the traditional carboxylates and phosphonates. We first present the synthesis of *n*-hexyl, 2-ethylhexyl, *n*-tetradecyl, *n*-octadecyl, and oleyl phosphinic acid. These compounds are suitable ligands for high-temperature nanocrystal synthesis (240-300°C) since, in contrast to phosphonic acids, they do not form anhydride oligomers. Consequently, CdSe quantum dots synthesized with octadecylphosphinic acid are conveniently purified, and their UV-Vis spectrum is free from background scattering. The CdSe nanocrystals have a low polydispersity and a photoluminescence quantum yield up to 18%. Furthermore, we could synthesize CdSe and CdS nanorods using phosphinic acid ligands, with high shape purity. We conclude that the reactivity towards TOP-S and TOP-Se precursors decreases in the series: cadmium carboxylate > cadmium phosphinate > cadmium phosphonate. By introducing a third and intermediate class of surfactants, we enhance the versatility of surfactant-assisted syntheses.

Keywords

nanocrystals, quantum dots, nanorods, ligands, phosphinic acids, surface chemistry

Introduction

Colloidal nanocrystals (NCs) are composed of nanosized (< 100 nm) inorganic crystalline cores, capped with (in)organic ligands on the surface.¹⁻² The most common organic ligands used in nanocrystal synthesis are also surfactants, with long alkyl chains and a polar binding group. For very small nanocrystals (< 10 nm) the classical concepts of colloidal stability do not apply and such nanocrystals have an equilibrium solubility. This *colloidal solubility* is determined by the binding group and the alkyl chain of the surfactant, the nanocrystal core size and the solvent.³⁻⁸ Apart from providing solubility to the final nanocrystals, surfactants also play an important role in nanocrystal synthesis, regulating the nucleation and growth rates.⁹⁻¹¹ Finally, controlling the surface chemistry has proven key to many (if not all) nanocrystal applications.², 9

Despite the crucial role of ligands, it is striking that most colloidal synthesis methods use the same types of surfactants: oleic acid, oleylamine and octadecylphosphonic acid being the most common. While thiols are great ligands for metals,¹² and metal chalcogenides,¹³ they are not thermally stable. Indeed, they are often used as sulphur precursors.¹⁴ Metal oleates are stable metal precursors for quantum dot syntheses at relatively low temperatures (240 °C).¹⁵⁻¹⁶ At higher temperatures they often decompose to metals or metal oxides.¹⁷⁻¹⁸ Oleylamine is a typical L-type ligand and binds usually in a dynamic fashion.¹⁹ In very specific systems, oleylamine can be a tightly bound ligand.²⁰⁻²¹ Phosphonates and phosphates are among the strongest ligands available and easily displace carboxylic acids,^{13, 22-23} especially when designed as multidentate ligands on a scaffold.²⁴⁻²⁵ Phosphonic acids are also the ligands of choice for high temperature (> 300 °C) quantum dot syntheses. Peng *et al.* used a mixture of saturated phosphonic acids to synthesize CdE (E = S, Se or Te) NCs and nanorods.²⁶ Later, wurtzite CdSe quantum dots were made by Owen *et al* using only *n*-octadecylphosphonic acid.²⁷ Carbone *et al.* used this strategy to obtain CdSe/CdS dot-in-rod structures.²⁸ Motivated by these reports, we recently developed

the synthesis of a library of phosphonic acid ligands,²³ including the highly soluble oleylphosphonic acid. Zhang *et al.* then used oleylphosphonic acid to synthesize CsPbBr₃ nanocrystals.²⁹⁻³⁰ Although phosphonic acids thus allow to make a variety of nanocrystal (hetero)structures, they are not without a drawback. We find that phosphonic acids are unstable at high temperature (300 °C), dehydrating to phosphonic acid anhydrides within 30 min (Figure S1). In nanocrystal syntheses with phosphonic acid ligands, phosphonic acid anhydrides are indeed often formed as by-products, and even retrieved on the nanocrystal surface.²⁹⁻³⁵ The formal dehydration is shown in Scheme 1. Many nanocrystal syntheses contain dehydration agents (e.g., phosphine chalcogenide or acyl bromide), thus forming other co-products than water. Since phosphonic acids have two OH moieties, this reaction can proceed further, forming oligomers.³² Over time, a macroscopic gel is often observed in nanocrystal samples with the phosphonic acid anhydride oligomer present.³¹ These may be a reason why reproducing certain nanocrystal reactions can prove challenging.

$$(n+2) \begin{array}{c} O \\ \parallel \\ P \\ OH \end{array} \xrightarrow{\Delta} HO \xrightarrow{||}{P} O \\ || \\ P \\ O \\ || \\$$

Scheme 1. Formal dehydration of phosphonic acids. In practice, often a dehydration agent is also present in the reaction mixture.

Another origin of irreproducibility is the use of impure reagents. In 2008, Wang *et al.* discovered the role of di-*n*-octylphosphinic acid and *n*-octylphosphinic acid on the formation of CdSe nanorods and nanowires.³⁶⁻³⁸ These molecules were present as impurities in commercially available tri-*n*-octylphosphine oxide (TOPO). Phosphinic acids were thus clearly an alternative ligand class, but only the di-substituted variants received further attention.³⁹ For example, bis(2,2,4-trimethylpentyl)phosphinic acid phosphinic acid was explored as co-ligand by Mulvaney *et al.*⁴⁰⁻⁴² Recently, Kovalenko *et al.* synthesized CsPbX₃ from PbBr₂, didodecyldimethylammonium halide and cesium di-isooctylphosphinate.⁴³ The di-alkyl

phosphinate did not end up as a ligand on the nanocrystal surface, presumably due to its high steric hindrance.⁴⁴⁻⁴⁵ Mono-substituted alkylphosphinic acids have not yet been explored in nanocrystal synthesis.

Here, we put forward mono-alkyl phosphinic acids as an alternative ligand class, similar to carboxylic acids and phosphonic acids. After presenting the ligand synthesis for several selected substrates, we proceed to show the intermediate reactivity of the phosphinic acids in CdSe quantum dot syntheses. The nanocrystals synthesized with phosphinic acids are also easier to purify since there is no gel formation. Very small (2-3 nm) CdSe quantum dots with low polydispersity and high photoluminescence quantum yields can be easily accessed with phosphinic acids ligands. CdSe and CdS nanorods were also synthesized using phosphinic acids, whereby the rods showed high purity and uniformity. We have thus presented a ligand class that is truly a worthy alternative to carboxylic acids and phosphonic acids, as X-type ligands in nanocrystal synthesis.

Results and Discussion

Phosphinic acid synthesis. To synthesize mono-alkyl phosphinic acids, suitable as ligands for nanocrystal synthesis, we identified two main methods.⁴⁶ Method I involves the addition of an alkene to hypophosphorous acid (H₃PO₂) or sodium hypophosphite (NaH₂PO₂). Double alkylation can occur and is typically avoided by an excess of phosphorus reagent. Several variations exist with different radical initiators, solvents and reagents concentrations.^{36, 46-49} We found the general protocol of Montchamp *et al.* (Scheme 2) most convenient to synthesize our target compounds; *n*-hexyl-, *n*-tetradecyl-, and *n*-octadecylphosphinic acid. The method can be easily performed on multigram scale and uses inexpensive reagents. We recrystallized the products to a high purity (>99%, based on ³¹P NMR).



Scheme 2. Method I to synthesize mono *n*-alkyl phosphinic acids and the obtained products with the yield indicated.

We also targeted more complex phosphinic acids such as oleylphosphinic acid and 2ethylhexylphosphinic acid. The oleyl fragment is ubiquitous in nanocrystals synthesis due to its high solubility and the high colloidal solubility it provides to nanocrystals. A branched chain provides also very high colloidal solubility as shown by Peng et al.³ To synthesize oleylphosphinic acid we first prepared the corresponding terminal alkene, since it is not commercially available. We preferred to start by reducing oleic acid to olevl alcohol to avoid the *trans* isomer present in technical oleyl alcohol. The reduction is performed with Li[AlH₄] according to Grela *et al.*⁵⁰ The alcohol was subsequently transformed in either a mesylate or a bromide to function as a good leaving group during the E_1 elimination towards the terminal alkene.⁵¹⁻⁵² Upon addition of KO^tBu, oleyl mesylate remained stable but oleyl bromide fully converted to the terminal alkene. Unfortunately, method I vielded a mixture of the desired linear oleylphosphinic acid and the undesired, branched octadec-1-en-9-ylphosphinic acid. For this reason, we used method II, where oleylbromide is converted into a Grignard reagent, and the latter reacts with diethyl chlorophosphite (Scheme 3).^{46, 53} Both the oleyl and the branched compound cannot be recrystallized and are purified (>95% according ³¹P NMR) by column chromatography as the ethyl ester, before being hydrolysed to the target product.



Scheme 3. Method II to synthesize more complex phosphinic acids and the obtained products. Conditions: (i) overnight at 50 °C in dry THF, then 2 h at room temperature in conc. HCl and water. (ii) 1.15 eq of TMS-Br, overnight in dry DCM, then dry MeOH, 6h, 40 °C.

CdSe quantum dot synthesis. Wurtzite CdSe quantum dots are typically synthesized at 300 °C from cadmium *n*-alkylphosphonate, which is formed *in situ* from cadmium oxide and *n*-alkylphosphonic acid. Oligomeric phosphonic acid anhydride is a co-product of this reaction, and it was observed previously that the final quantum dot suspension turns in a macroscopic gel over time.³¹ Drijvers *et al.* managed to significantly reduce the formation of the oligomeric phosphonic acid anhydride upon addition of oleyl alcohol to the reaction.³¹ The alcohol forms an ester with one of the two hydroxyl groups of the phosphonic acid, thus inhibiting oligomer formation. Since also gel formation was thus inhibited, it was suggested that the gel formation is related to the presence of phosphonic acid anhydride oligomer, although the exact mechanism of gelation is unknown. We hypothesize that we avoid the problem entirely by using mono-alkyl phosphinic acids.

Similar to the phosphonic acid based strategy,²⁶⁻²⁷ we first dissolved CdO with two equivalents of octadecylphosphinic acid in TOPO. Phosphinic acid requires higher temperatures to dissolve the CdO, compared to phosphonic acid. This is most likely due to the lower acidity of alkylphosphinic acid (pKa in H₂O \approx 3.1) compared to alkylphosphonic acid (pKa in H₂O \approx 2.4 and 7.7).⁵⁴ After dissolution of CdO, TOP-Se is injected at 300 °C and CdSe quantum dots are rapidly formed (Figure 1). After 10 minutes the reaction mixture is cooled down and the nanocrystals are easily purified by precipitation/redispersion cycles with acetone/toluene as

antisolvent/solvent. We did not observe any oils or gels. Although the nanocrystals have already ripened (see next section), the size dispersion is still quite low; 8 % (size according to TEM: 3.46 ± 0.28 nm ($\mu \pm \sigma$)), see Figure 1B and Figure S2. Given the small nanocrystal size, we turned to x-ray Pair Distribution Function analysis (PDF) to determine the crystal structure.⁵⁵ The refinement using the wurtzite crystal structure gave a much better fit ($R_w = 0.17$) than the refinement with the zincblende crystal structure ($R_w = 0.50$), see Figure 1C and Figure S3. From the wurtzite refinement, we determined the crystallite size to be 2.9 nm (Figure S3). The nanocrystal dispersion is also free from organic impurities as attested by NMR. In the ¹H NMR spectrum we observe the broadened signature of a bound alkyl chain (Figure 1D).⁵⁶ In Diffusion Ordered Spectroscopy (DOSY), we find only a single diffusing species (apart from toluene), see Figure 1E. The diffusion coefficient is 93.78 ± 0.16 µm² s⁻¹, corresponding to a solvodynamic diameter of 7.8 nm (which agrees quite well with the nanocrystal core size + twice the length of the octadecyl chain (approx. 2 nm). There is no evidence of an oligomeric impurity while we find this impurity in the DOSY spectrum of CdSe quantum dots synthesized with octadecylphosphonic acid (Figure S4).



Figure 1. CdSe quantum dots synthesized with *n*-octadecylphosphinic acid. (A) general reaction scheme, (B) TEM image with histogram, (C) x-ray PDF analysis showing a single phase refinement with the wurtzite crystal structure. (D) ¹H NMR spectrum, and (E) DOSY NMR spectrum.

In the ³¹P NMR spectrum, we observe a broadened resonance around 25 ppm, assigned to bound ligands (Figure S5A). In the ¹H NMR spectrum, the ratio of the methylene (-CH₂-) and methyl (CH₃-) integral (3:30) is lower than expected for an octadecyl chain (3:34), see Figure S5A. This suggests that a shorter chain is present. After stripping the native ligands with trifluoroacetic acid, we do not observe the expected P-H resonance of *n*-alkylphosphinic acid in the ¹H NMR spectrum (Figure S5B). Also in ³¹P NMR spectrum, the resonance of the stripped ligand does not agree with the reference spectrum of alkylphosphinic acid. It does match with the reference spectrum of alkylphosphinic acid. These result suggests that during the synthesis a portion of the octadecylphosphinic acids oxidized and that the solvent (TOPO) decomposed and formed octylphosphonic acid. There is literature precedent for the latter.³⁴⁻³⁵ Alternatively, we stripped the native ligands with chlorotrimethylsilane, and we observe 2

major resonances matching with trimethylsilyl hydrogen phosphonate and bis(trimethylsilyl) phosphonate (Figure S5C).²⁹ Quenching the stripping products with anhydrous methanol results in a single resonance matching with alkylphosphonic acid (Figure S5C). In addition, MS analysis of these products is consistent with the presence of two compounds: octadecylphosphonic acid (major component) and octylphosphonic acid (minor component), see Figure S6.

Kinetic comparison with phosphonic and carboxylic acid ligands. According to UV-Vis spectroscopy, CdSe quantum dots synthesized with phosphonic acid (5.4 nm) are larger than the ones synthesized with phosphinic acids (3.3 nm), see Figure S7. This could be related to the precursor conversion kinetics of the reactions since it is established that faster reactions yield more, but smaller, particles.^{15, 57} To investigate this in more detail, we followed the reaction kinetics and also compared with carboxylic acid ligands (Figure 2). We took several reaction aliquots, quantified and diluted them for quantitative UV-Vis spectroscopy analysis. In case of the mono-alkyl phosphinic acids, the kinetic traces showed a good batch-to-batch reproducibility, see Figure S8. The nanocrystal size was calculated from the position of the first excitonic peak, and the absorption at 340 nm was used to determine the yield.⁵⁸ From the yield and the size, we calculate the number of particles. We performed this analysis at 300 °C for both phosphinic and phosphonic acid ligands, and at 240 °C for both phosphinic and carboxylic acid ligands. These temperatures were chosen since 300 °C is the common reaction temperature for phosphonic acid ligands, and 240 °C is usual for carboxylic acids.

For phosphonic acids, the yield shows comparable kinetics to the previous reports for the used Cd:Se stoichiometry (Figure 2).³³ After an induction time (where no particles are observed), the number of particles increases over the course of the reaction suggesting continuous nucleation instead of burst nucleation. Continuous nucleation was recently found to be a quite general feature of nanocrystal synthesis, applicable to Ir, Pd, InP and CdSe nanocrystals.^{10, 59-61} The

particles grow steadily from 3.4 to 5.4 nm, while the valley-to-peak ratio slightly increases. Given the difficulty in calculating an exact polydispersity from UV-Vis spectroscopy, we choose to present the valley-to-peak ratio, as a measure for the polydispersity at a given size.⁶² A low valley-to-peak ratio is characteristic for a monodisperse sample, but for a constant polydispersity, the valley-to-peak ratio increases with nanocrystal size. As such, we do not interpret the slight increase in valley-to-peak ratio over the course of the reaction as ripening.



Figure 2. UV-Vis absorption spectra of CdSe quantum dots synthesized from a CdX_2 salt and TOP-Se, at two temperatures, with X being either phosphonate, phosphinate, or carboxylate. Besides the used ligand and temperature, all other reaction parameters remained identical. Also the extracted mean diameter, the valley-to-peak ratio, the reaction yield and the number of particles are plotted for the four different reactions. Note that the scale changes for the number of particles across the four figures.

With phosphinic acids we did not observe an induction time. At 300 °C, particles are formed almost instantaneously and the first particles are also much smaller (2.5 nm) than the first particles observed for phosphonic acids (3.4 nm). The reaction reaches also a higher yield after 10 min (72 % vs 28 % for phosphinic and phosphonic acids respectively). Most of the reaction with phosphinic acids is already completed after 2 min. However, the particles continue to grow, presumably through a ripening mechanism since the number of particles slightly

decreases and the valley-to-ratio significantly increases (Figure 2). Clearly, phosphinic acids allow for a higher reactivity than phosphonic acids (faster yield development). Furthermore, the formed particles with phosphinic acids are particularly small (< 3 nm, a size range that is usually difficult to access⁶³) and the particles have sharp first excitonic features (FWHM at 2.7 eV: 260 meV).⁶⁴⁻⁶⁵ An additional advantage of the phosphinic acids for kinetic analysis is the long term stability of the dispersions. While kinetic analysis has to be performed immediately for the phosphonic acids samples due to partial gelation (causing scattering in UV-Vis spectroscopy), the phosphinic acid samples give identical results after 20 weeks, see Figure S9.

At 240 °C (Figure 2), the yield development seems more sluggish for phosphinic acids compared to carboxylic acids, suggesting that cadmium phosphinates are less reactive than cadmium carboxylates. Note that we could not determine the yield (and thus the particle number) for the aliquots taken until 2 min since the absorption at 340 nm is not size independent for these very small sizes (< 2.7 nm). For carboxylic acids, we see again continuous nucleation (increasing particle number as the yield increases, see Figure 2), while we do not observe such a feature for the phosphinic acids. The particle number remains constant while the yield increases slightly. We do not observe ripening for the phosphinic acids at 240°C. Instead, the size dispersion focuses (indicated by a decreasing valley-to-peak ratio) while the particles are growing. Again, very small particles (2-3 nm) are obtained with sharp excitonic features (FWHM at 2.54 eV: 244 meV). We conclude that the phosphinic acid ligands lead to particle ensembles with narrow size dispersions and that their reactivity is intermediate between phosphonic and carboxylic acids.

Photoluminescence properties. For crude CdSe (i.e., unpurified reaction mixture) synthesized with oleic acid at 240 °C, we obtained a photoluminescence quantum yield (PLQY) of 5.6% (Figure S10A). After purification, this decreased to 0.5%, an effect which has been observed before in the literature.^{64, 66-68} For the CdSe particles synthesized with phosphonic acids, we

obtained a very low PLQY of 0.2% for the crude and < 0.1% after purification (Figure S10B). Concerning CdSe synthesized with phosphinic acid under identical conditions at 300 °C, we obtained a PLQY of 18% for the crude aliquot at 10 seconds (first excitonic peak at 2.7 eV), with a FHWM of 150 meV (Figure S10C). After ripening (10 min), the PLQY is 16.1 \pm 0.8 % (average and error determined from four independent syntheses) and after purification we measure only 3%. For CdSe synthesized with phosphinic acid at 240 °C, there is more variability and we measure a PLQY of 6-11% for the crude after 15 min reaction (first excitonic peak at 2.53 eV) and the FWHM was 130 meV (Figure S10D). After purification, the PLQY reduced to 4%. A full photophysical analysis is outside the scope of this work, but it is clear that under identical conditions, CdSe nanocrystal synthesized with phosphinic acids. Their values are up to par with the state of the art of unshelled CdSe nanocrystals (typically less than 15%, with the exception of Bawendi *et al.*⁶⁹ and Jasieniak *et al.*⁷⁰), and with quite narrow line-widths.^{66, 68-77}

Synthesis of CdSe and CdS nanorods. Phosphonic acids are also used in reactions to obtain anisotropic nanocrystals, such as nanorods. Buhro *et al.* synthesized CdSe nanorods by reacting cadmium tetradecylphosphonate with TOP-Se in the presence of di-*n*-octylphosphinic acid (DOPA).³⁶ The TOP-Se is injected at 275 °C but the temperature is allowed to drop to 250 °C during the growth of the rods. The amount of added DOPA controls the aspect ratio. We successfully reproduced this method but we obtained, apart from the intended rods, also tetrapods (Figure 3 and Figure S11). When we perform the same reaction with cadmium tetradecylphosphinate, no rods but regular quantum dots are obtained (Figure S12). The reaction reaches maximum yield at 1 min, after which the particles start ripening. We attribute this result to the higher reactivity of cadmium phosphinate, depleting all the precursor already during the formation of the seeds. Therefore, we lowered also the injection temperature to 250 °C, and particles with a slightly anisotropic shape are obtained (Figure S13). Decreasing the temperature even further (TOP-Se is injected at 250 °C and the temperature is allowed to drop to 225 °C for the growth), results in nanorods albeit with a small aspect ratio and some tetrapods are again observed, see Figure 3 and Figure S14.



Figure 3. CdS and CdSe nanorods synthesized with phosphonic and phosphinic acid.

TOP-S has a higher phosphor-chalcogen bond dissociation energy (BDE) than TOP-Se.¹⁸ We thus hypothesized that cadmium phosphinate would be more suitable to grown CdS rods since the higher BDE of TOP-S would decrease the precursor conversion rate. Gratifyingly, CdS rods were indeed obtained when TOP-S was injected at 275 °C and the rods were grown at 250 °C (Figure 3). The nanorods were highly monodisperse (width: 2.4 ± 0.3 nm, and length: 14.6 ± 2.8 nm) and the sample had >99% shape purity (Figure S15). In comparison, the control experiment with phosphonic acid failed several times. In one case the reaction resulted in CdS rods but they were contaminated with tetrapods (Figure 3). Furthermore, the rods with phosphinic acid were easier to purify due to the absence of a gel.

Discussion. We have introduced another surfactant class: mono-alkylphosphinic acids. Several features make this ligand class stand out. (1) Both the kinetics experiments and the nanorod synthesis have confirmed that cadmium phosphinates have intermediate reactivity, reacting slower with TOP-E precursors than cadmium carboxylates but faster than cadmium phosphonates. This provides the nanochemist with an additional handle on the reaction kinetics.

Before, tuning the precursor conversion kinetics was limited to the chalcogen or pnictogen precursor,^{16, 59, 78-80} but now we gain an additional handle on the kinetics through the metal precursor salt. (2) Reactions with phosphinic acids are not plagued by the occurrence of oligo-anhydride. The latter can manifest itself by inducing macroscopic gel formation, or as background scattering in UV-Vis spectroscopy or SAXS (small-angle x-ray scattering). Therefore, we expect that this class of mono-alkyl phosphinic acids will help to further improve the quantitative analysis of quantum dots (sizing curves and extinction coefficients). (3) CdSe quantum dots synthesized with phosphinic acids fall into a size regime (2-3 nm) that is traditionally difficult to access, especially for the wurtzite crystal structure.⁶³ The particles are monodisperse and have a high photoluminescence quantum yield.

Although we have used here the cadmium chalcogenides as our model system to showcase the versatility of mono-alkyl phosphinic acids, this ligand class is expected to be more widely applicable in nanocrystal science. These ligands were indeed not designed for a specific material. For example, unlike thiol ligands, they are expected to have also a good binding affinity to metal oxide nanocrystals (e.g., ZrO₂), but this remains to be tested. Other open questions concern the nucleation mechanism. It is striking that we easily found supporting evidence for continuous nucleation in case of phosphonic acids and carboxylic acids but none for phosphinic acids. Given that mono-alkyl phosphinic acids seems more well-behaved than alkylphosphonic acids, mechanistic studies are more convenient and could tackle these open questions.

Conclusion

We have presented convenient strategies to synthesize phosphinic acids that are relevant for nanocrystals synthesis. Furthermore, cadmium phosphinate was shown to react with TOP-Se to form CdSe nanocrystals of high purity, low polydispersity and reasonably high photoluminescence quantum yield. We also demonstrated the synthesis of CdSe and CdS nanorods with high shape uniformity for the latter. Compared to nanocrystals synthesized with phosphonic acids, the ones synthesized with phosphinic acids are more easily purified since no gel is formed. Interestingly, the reactivity of cadmium phosphinate is intermediate between cadmium phosphonate and cadmium carboxylate. This provides researchers with an additional handle on the kinetics of precursor conversion, which seems so far dominated by tuning the chalcogen precursors. In addition, we did not find any indication of continuous nucleation in the case of cadmium phosphinate. While this points to interesting mechanistic differences, more detailed studies are necessary to uncover its wider implications.

Experimental

General considerations. All manipulations are performed in air, unless otherwise indicated. All chemicals are used as received unless otherwise mentioned. *1*-Hexene (99%), hypophosphorous acid (50% aqueous solution), lithium aluminium hydride (95%), potassium hydrogen sulfate (99%), and triethylborane (1M in THF) were purchased from Acros. *1*-Tetradecene (94%), cadmium oxide (99.998%), calcium hydride (92%), benzene (99.5%), magnesium turnings (99.9%), magnesium sulfate (99.0%), sulfur (99.5%), and chloroform-d1 (99.8%D stabilized with Ag) were purchased from Carl Roth. Acetone (99%), acetonitrile (99.9%), celite, chloroform (99.5%), dichloromethane (99.8%), diethyl ether (99.5%), dioxane (99.9%), ethanol absolute (99.9%), ethyl acetate (99.5%), *n*-heptane (99%), *n*-hexane (99%), hydrochloric acid (37%), methanol (99%), silica gel 60A (40-63 μm), sodium chloride (99.9%), sodium hydroxide (98.5%), sodium sulfate (99%), tetrahydrofuran (99.9%), toluene (99.9%), and trifluoroacetic acid (99%) were purchased from Chem-Lab. *n*-Hexylphosphonic acid (97%), *n*-tetradecylphosphonic acid (97%), and *n*-octadecylphosphonic acid (97%) were purchased from PlasmaChem. 1,2-Dibromoethane (98%), 1-bromooctadecane (97%), 1octadecene (90%), 1-octene (98%), 2-ethyl-hexylbromide (95%), benzoylperoxide (moistened with 25% water). bromotrimethylsilane (97%), diethyl chlorophosphite (95%), methanesulfonyl chloride (99.7%), oleic acid (90%), potassium carbonate (99%), potassium hydroxide (90%), potassium *tert*-butoxide (98%), sodium hypophosphite monohydrate (99%) tetrabromomethane (99%), trimethylamine (99%), and triphenylphosphine (99%) were purchased from Sigma-Aldrich. Selenium (99.99%), tri-n-octylphosphine (90%), and tri-noctylphosphine oxide (90%) were purchased from STREM. Trifluoroacetic anhydride was purchased from TCI. Benzene-d₆ (99.5%D), methanol- d_4 (99.8%D), and toluene-d₈ (99.5%D) were purchased from VWR.

Some of the chemicals are toxic (*e.g.*, cadmium oxide), carcinogenic (*e.g.*, dichloromethane),⁸¹ or pyrophoric (*e.g.*, lithium aluminium hydride),⁸² and can have a severe impact on health and environment on short and/or long term. It is advised to consult the available Safety Data Sheets (SDS) provided by the producer prior to use. Specific safety consideration for NC synthesis making use of TOPO and TOP have been described by Schaak *et al.* in more detail.⁸³

When required, organic solvents are dried according to the procedure described by Williams *et* $al.^{84}$ making use of 20% m/v freshly activated 3Å sieves for minimum 120 hours.

Trioctylphosphine oxide (TOPO) is recrystallized on a scale of 200 g by the procedure described by Owen *et al.* to obtain purified white needles (178 g, 89 %),^{27, 37} resulting in the same structure obtained by previous single-crystal XRD measurements.⁸⁵ Sulfur is recrystallized on a scale of 12.5 g by the following procedure. Sulfur is dissolved in toluene (1 mg/mL) at 100 °C and filtered over a hot frit. While the solution is cooling down to room temperature, shiny yellow needles grow. Afterwards, these needles are filtered and extensively

vacuum dried (9.0 g, 72 %). *n*-Octadecylphosphonic acid is synthesized according to the procedure described by De Roo *et al.* on a 75 mmol *n*-octadecyl bromide scale (18.9 g, 76 %).²³ Di-*n*-octylphosphinic acid is synthesized according to the procedure described by Buhro *et al.* on a 200 mmol hypophosphorous acid scale (17.2 g, 30 %).³⁷

Synthesis of mono-alkylphosphinic acid. Mono-alkylphosphinic acids are synthesized according to the procedure of Montchamp et al. with slight adaptations.⁴⁸ An open 250 mL flask is loaded with sodium hypophosphite hydrate (6.63 g, 62.5 mmol, 2.5 eq.), 1-alkene (25 mmol, 1.0 eq.), methanol (125 mL), and 1 mol L⁻¹ triethylborane in tetrahydrofuran (25 mL, 25 mmol, 1.0 eq.) which is vigorously stirred for 3 hours open to the air. Afterwards, the reaction mixture is transferred to a separating funnel (1 L) to which dichloromethane (125 mL) and 1 mol L⁻¹ solution of KHSO₄ (125 mL) is added. The aqueous phase is washed once again with dichloromethane (125 mL). All the organic phases are collected, dried with Na₂SO₄, and concentrated on the rotary evaporator. For solid products, such as n-octadecyl- and ntetradecylphosphinic acid, a recrystallization from hot *n*-hexane (25 mL) is performed twice to obtain a final purified white product. For liquid products, such as *n*-hexylphosphinic acid, the product is first transformed to its potassium salt by adding a stoichiometric amount of potassium hydroxide in EtOH (0.5 mol L⁻¹). After 1 hour stirring, the salt is concentrated on the rotary evaporator and twice recrystallized from a hot mixture of *n*-hexane (10 mL), tetrahydrofuran (10 mL), and ethanol (1 mL). The mixture is slowly cooled down to room temperature, fridge, and overnight in the freezer (-20 °C). The precipitated product is filtered and collected. Purity is checked using ³¹P NMR (if a residual peak at 3 ppm of hypophosphorous salt is observed, no additional recrystallization is needed since it will get removed in the next washing step). To obtain the final purified *n*-hexylphosphinic acid, the purified potassium *n*-hexylphosphinate salt is acidified with conc. HCl (2 mL) in H₂O (5 ml) and extracted three times with dichloromethane (5 mL). The organic fraction is collected and dried with Na₂SO₄, concentrated on the rotary evaporator, and extensively vacuum dried. This reaction can be run at four times the scale described with no observable changes.

n-Octadecylphosphinic acid (5.1 g, 61.0 %). ¹**H NMR** (300MHz, CDCl₃): δ 8.70 (s, 1H), 8.00 (t, *J* = 1.8 Hz, 0.5H), 6.20 (t, *J* = 1.8 Hz, 0.5H), 1.75 (quin, *J* = 8.5 Hz, 2H), 1.65-1.45 (m, 2H), 1.45-0.95 (m, 30H), 0.85 (t, *J* = 7.0 Hz, 3H). ¹³**C NMR** (75MHz, CDCl₃): δ 32.10, 30.67, 30.50, 30.24, 29.87, 29.75, 29.53, 29.30, 28.99, 22.85, 20.62, 14.28. ³¹**P**[1H] NMR (75MHz, CDCl₃): δ 40.14 ppm. **LC-MS** (API-ES) calc for C₁₈H₃₈O₂P [M-H]⁻ 317.26, found 317.2.

n-*Tetradecylphosphinic acid* (3.2 g, 48.1 %). ¹**H NMR** (300MHz, CDCl₃): δ 9.45 (s, 1H), 8.00 (t, *J* = 1.8 Hz, 0.5H), 6.20 (t, *J* = 1.8 Hz, 0.5H), 1.75 (quin, *J* = 8.5 Hz, 2H), 1.60-1.35 (m, 2H), 1.35-0.95 (m, 22H), 0.85 (t, *J* = 7.0 Hz, 3H). ¹³**C NMR** (75MHz, CDCl₃): δ 31.92, 30.52, 30.31, 30.02, 29.69, 29.66, 29.63, 29.59, 29.36, 29.13, 28.78, 22.69, 20.45, 14.11. ³¹P[1H] NMR (75MHz, CDCl₃): δ 39.6 ppm. **LC-MS** (API-ES) calc for C₁₄H₃₂O₂P [M+H]⁺ 263.21, found 263.2.

Potassium n-hexylphosphinate (3.0 g, 63.2 %). ¹H NMR (300MHz, MeOD-d₄): δ 7.78 (t, J = 1.6 Hz, 0.5H), 6.16 (t, J = 1.6 Hz, 0.5H), 1.65-1.05 (m, 10H), 0.91 (t, J = 6.9 Hz, 3H). ¹³C NMR (75MHz, MeOD-d₄): δ 34.37, 32.75, 31.79, 23.59, 23.05, 14.42. ³¹P[1H] NMR (75MHz, MeOD-d₄): δ 27.8 ppm. LC-MS (API-ES) calc for C₆H₁₀O₂PK [M-K]⁻ 149.07, found 149.2.

n-Hexylphosphinic acid (1.6 g, 42.0 %). ¹**H NMR** (300MHz, CDCl₃): δ 9.45 (s, 1H), 8.00 (t, J = 1.8 Hz, 0.5H), 6.20 (t, J = 1.8 Hz, 0.5H), 1.75 (quin, J = 8.5 Hz, 2H), 1.65-1.45 (m, 2H), 1.35-0.95 (m, 6H), 0.85 (t, J = 7.0 Hz, 3H). ¹³**C NMR** (75MHz, CDCl₃): δ .31.40, 30.07, 28.64, 22.48, 20.67, 14.09. ³¹**P**[1H] NMR (75MHz, CDCl₃): δ 38.7 ppm. **LC-MS** (API-ES) calc for C₆H₁₆O₂P [M+H]⁺ 151.09, found 151.1.

Synthesis of (Z)-octadec-9-en-1-ol (oleyl alcohol). Oleyl alcohol is synthesized by reducing oleic acid according to the procedure published by Grela *et al.*⁵⁰ In a 1 L flask, LiAlH₄ (10 g,

260 mmol, 3.5 eq.) is added, degassed three times, and put under argon. After the addition of dry THF (300 mL), the flask is cooled to 0 °C in an ice bath. Dried oleic acid (21.48 g, 24.0 mL, 76 mmol, 1.0 eq.) is added dropwise. The reaction is allowed to heat up to room temperature and stirred for 4 hours. Afterwards, the reaction flask is again cooled to 0 °C just before diethyl ether (400 mL), H₂O (10 mL), 15% NaOH solution (10 mL), and H₂O (32 mL) is carefully added. After 15 minutes of additional stirring, the reaction mixture is dried using MgSO₄, and filtered over celite. The filtrate is first vacuum dried, and then vacuum distilled following the same vacuum distillation procedure as described above to obtain a colourless viscous liquid (17.5 g, 86 %). ¹H NMR (400MHz, CDCl₃): δ 5.40-5.25 (m, 2H), 3.60 (t, *J* = 6.6 Hz, 2H), 2.05-1.90 (m, 4H), 1.55 (quin, *J* = 7.0 Hz, 2H), 1.45 (s, 1H), 1.40-1.10 (m, 22H), 0.85 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (75MHz, CDCl₃): δ 130.09, 129.94, 63.18, 32.93, 32.04, 29.90, 29.88, 29.66, 29.64, 29.55, 29.46, 29.37, 27.35, 27.33, 25.88, 22.82, 14.24. LC-MS (API-ES) calc for C₁₈H₃₇O [M+H]⁺ 269.28, found 269.3.

Synthesis of (*Z*)-octadec-9-en-*1*-yl methane sulfonate (oleyl mesylate). In a 250 mL Schlenk flask, oleyl alcohol (13.42 g, 15.51 mL, 50 mmol, 1.0 eq.) is added, degassed three times, and put under argon. After the addition of dry dichloromethane (100 mL) and dried triethylamine (5.57 g, 7.67 mL, 55 mmol, 1.1 eq.), the flask is cooled to 0 °C in an ice bath. Dry mesylate chloride (6.30 g, 4.26 mL, 55 mmol, 1.1 eq.) is added dropwise and further stirred for 30 minutes, followed by the addition of H₂O (50 mL). After 30 minutes of additional stirring, the reaction mixture is transferred to a separation funnel (300 mL) where the organic phase is kept and the aqueous phase is extracted twice with chloroform (50 mL). All the organic phases are collected together and washed with: (1) a mixture of 25% K₂CO₃ solution (25 mL) and H₂O (50 mL); (2) brine (50 mL), and dried with MgSO₄. After vacuum drying, a colourless viscous liquid is obtained (17.2 g, 99 %). ¹H NMR (300MHz, CDCl₃): δ 5.40-5.25 (m, 2H), 4.20 (t, *J* = 6.6 Hz, 2H), 2.95 (s, 3H), 2.10-1.95 (m, 4H), 1.70 (quin, *J* = 7.5 Hz, 2H), 1.50-1.10 (m, 22H),

0.85 (t, J = 7.2 Hz, 3H). ¹³C NMR (75MHz, CDCl₃): δ 130.18, 129.86, 70.30, 37.51, 32.05, 29.90, 29.83, 29.66, 29.46, 29.28, 29.15, 27.36, 27.30, 25.56, 22.82, 14.25. LC-MS (API-ES) calc for C₁₉H₄₂NO₃S [M+NH₄]⁺ 364.29, found 364.3.

Synthesis of (*Z*)-octadec-9-en-1-yl bromide (oleyl bromide). Oleyl bromide is synthesized according to the procedure of De Roo *et al.* with slight adaptations.²³ A 250 mL flask is loaded with oleyl alcohol (13.42 g, 15.81 mL, 50 mmol, 1.0 eq.), tetrabromomethane (18.24 g, 55 mmol, 1.1 eq.), and dry dichloromethane (50 mL), and cooled to 0 °C with an ice bath. Triphenylphsophine (15.74 g, 60 mmol, 1.2 eq.) was added in portions over 15 minutes. The reaction was allowed to stir for 1 hour at room temperature. Afterwards, hexanes (50 mL) is added to the mixture, cooled in the freezer to precipitate triphenylphosphine oxide, filtered over a silica plug, and the filtrate is evaporated. This step is repeated a second time to remove all the triphenylphosphine oxide, resulting in a colourless liquid (16.3 g, 98 %) which is vacuum dried overnight to remove bromoform, and stored in fridge. ¹H NMR (300MHz, CDCl₃): δ 5.40-5.25 (m, 2H), 3.40 (t, *J* = 6.9 Hz, 2H), 2.10-1.90 (m, 4H), 1.85 (quin, *J* = 7.7 Hz, 2H), 1.50-1.10 (m, 22H), 0.85 (t, *J* = 7.0 Hz, 3H). ¹³C NMR (75MHz, CDCl₃): δ 130.16, 129.91, 34.13, 33.01, 32.07, 29.94, 29.88, 29.70, 29.49, 29.35, 28.92, 28.35, 27.40, 27.33, 22.86, 14.28. GC-MS molecular mass for C₁₈H₃₅Br [M] 331.38, found 331 (average mass).

Synthesis of (Z)-octadeca-1,9-diene. A 500 mL flask is loaded with oleyl bromide (16.57 g, 50 mmol, 1.0 eq.), evacuated, and set under argon atmosphere. Dry tetrahydrofuran (200 mL) is added and cooled down to 0 °C with an ice bath. Potassium *tert*-butoxide (1M in THF, 115 mL, 115 mmol, 2.3 eq.) is added dropwise. It is important that a full yield is obtained since separation between the diene and bromide is not straightforward. The reaction mixture is allowed to warm to room temperature after the addition of the first equivalent of base and stirred overnight. Afterwards, chloroform (200 mL) is added, and the mixture is washed twice with water (200 mL) which is discarded. The organic phase is evaporated, and vacuum distilled over

CaH₂ to result in a colourless liquid (9.4 g, 75 %).¹**H NMR** (400MHz, CDCl₃): δ 5.90-5.70 (m, 1H), 5.40-5.25 (m, 2H), 5.05-4.85 (m, 2H), 2.10-1.90 (m, 6H), 1.50-1.15 (m, 20H), 0.85 (t, *J* = 7.1 Hz, 3H). ¹³**C NMR** (75MHz, CDCl₃): δ 139.34, 130.12, 129.95, 114.29, 33.97, 32.08, 29.95, 29.89, 29.70, 29.50, 29.32, 29.20, 29.08, 27.40, 27.36, 22.87, 14.26. **GC-MS** molecular mass for C₁₈H₃₄ [M] 250.47, found 250 (average mass).

Synthesis of ethyl (2-ethylhexyl)phosphinate ester according to the procedure of Meier et al.⁵³ with slight adaptations. In an argon filled glovebox, a 250 mL Schlenk flask is loaded with diethyl chlorophosphite (3913.8 mg, 3.595 mL, 25 mmol, 1.00 eq.) and dry tetrahydrofuran (50 mL). The flask is transferred to the Schlenk line and cooled down to 0 °C in an ice bath. Separately, an oven dried 100 mL Schlenk flask is loaded with activated (sanded) magnesium turnings (1.3 g, 52.5 mmol, 2.15 eq.), dry tetrahydrofuran (50 mL), 2-ethyl-hexylbromide (3478.0 mg, 3.205 mL, 25 mmol, 1.00 eq.), a few drops of dry 1,2-dibromoethane, and is stirred at room temperature overnight prior to a filtered Cannula transfer into the first solution. After the addition, the reaction mixture is stirred overnight at 50 °C. The next day, the solvent is evaporated, concentrated hydrochloric acid (5 mL) and water (75 mL) is added and stirred for 2 hours at room temperature. Afterwards, the reaction mixture is extracted twice with dichloromethane (75 mL), dried with Na₂SO₄, concentrated on the rotary evaporator, and purified using silica gel column chromatography (EtOAc). The final product is vacuum dried overnight to remove all solvent traces and result in a colourless liquid (2.5 g, 49 %). ¹H NMR $(400 \text{ MHz}, \text{ CDCl}_3)$: δ 7.75 (t, J = 1.9 Hz, 0.5 H), 6.45 (t, J = 2.1 Hz, 0.5 H), 4.20-3.95 (m, 2H), 1.85-1.65 (m, 3H), 1.50-1.10 (m, 11H), 0.90-0.75 (m, 6H). ¹³C NMR (100MHz, CDCl₃): δ 62.40, 33.89, 33.54, 32.65, 28.57, 27.14, 22.90, 16.34, 14.11, 10.57. ³¹P[1H] NMR (160MHz, CDCl₃): δ 38.53 ppm. LC-MS (API-ES) calc for C₁₀H₂₄O₂P [M+H]⁺ 207.15, found 207.2.

Synthesis of ethyl (oleyl)phosphinate ester is synthesized using the same procedure as for ethyl (2-ethylhexyl)phosphinate ester but with oleyl bromide (8284.3 mg, 25 mmol, 1.00 eq.)

instead of 2-ethyl-hexylbromide. The final product is a colourless liquid (1.6 g, 18.6 %). ¹**H NMR** (400MHz, CDCl₃): δ 7.70 (t, *J* = 1.8 Hz, 0.5H), 6.40 (t, *J* = 1.9 Hz, 0.5H), 5.40-5.25 (m, 2H), 4.25-3.95 (m, 2H), 2.10-1.85 (m, 4H), 1.80-1.65 (m, 2H), 1.65-1.50 (m, 2H), 1.50-1.15 (m, 25H), 0.85 (t, *J* = 6.8 Hz, 3H). ¹³**C NMR** (100MHz, CDCl₃): δ 130.12, 129.84, 62.38, 32.02, 30.63, 30.48, 29.89, 29.80, 29.65, 29.44, 29.37, 29.31, 29.24, 28.42, 27.37, 27.28, 22.79, 20.84, 16.40, 14.22. ³¹**P**[1**H**] **NMR** (160MHz, CDCl₃): δ 38.94 ppm. **LC-MS** (API-ES) calc for C₂₀H₄₂O₂**P** [M+H]⁺ 345.29, found 345.3.

Synthesis of 2-ethylhexyl and oleylphosphinic acid. The phosphinate is dissolved in dry dichloromethane (2 mL per gram phosphinate) in dried Schlenk flask. TMS-Br (1.15 equivalents) is added and the reaction mixture is stirred overnight under argon. The volatiles are removed by evaporation, excess dry methanol (5 mL per gram phosphinate) is added and the solution is stirred for 6 hours at 40 °C. Afterwards, the solvent is removed by evaporation and the product is vacuum dried overnight at 50 °C to remove methanol.

2-*Ethylhexylphosphinic acid, quantitative yield.* ¹**H NMR** (400MHz, CDCl₃): δ 11.50 (s, 1H) 7.85 (s, 0.5H), 6.50 (s, 0.5H), 1.90-1.65 (m, 3H), 1.55-1.10 (m, 8H), 0.95-0.75 (m, 6H). ¹³**C NMR** (100MHz, CDCl₃): δ 33.90, 33.81, 33.36, 28.53, 27.10, 22.93, 14.16, 10.52. ³¹P[1H] **NMR** (160MHz, CDCl₃): δ 38.29 ppm. **LC-MS** (API-ES) calc for C₈H₂₀O₂P [M+H]⁺ 179.12, found 179.2.

Oleylphosphinic acid, quantitative yield. ¹**H NMR** (400MHz, CDCl₃): δ 11.50 (s, 1H), 7.75 (s, 0.5H), 6.40 (s, 0.5H), 5.40-5.25 (m, 2H), 2.10-1.85 (m, 4H), 1.85-1.65 (m, 2H), 1.65-1.50 (m, 2H), 1.50-1.15 (m, 22H), 0.85 (t, *J* = 7.0 Hz, 3H). ¹³**C NMR** (100MHz, CDCl₃): δ 130.14, 129.88, 32.07, 30.64, 30.49, 29.93, 29.87, 29.68, 29.47, 29.42, 29.38, 29.26, 28.94, 27.38, 27.32, 22.83, 20.71, 14.27. ³¹**P**[1H] NMR (160MHz, CDCl₃): δ 38.83 ppm. **LC-MS** (API-ES) calc for C₁₈H₃₆O₂P [M-H]⁻ 315.25, found 135.2.

Synthesis of cadmium *n***-tetradecylphosphinate.** The procedure is inspired by the one used to synthesize cadmium oleate of Hendricks *et al.*^{15-16, 86} CdO (642.1 mg, 5 mmol, 1eq.) and acetonitrile (4 mL) are stirred at 0 °C in an ice bath. Trifluoroacetic anhydride (708 μ L, 5 mmol, 1 eq.) and trifluoroacetic acid (77 μ L, 1 mmol, 0.2 eq.) are added and allowed to warm to room temperature and results in a clear and colourless solution overnight, (if the resulting solution would have a white precipitate, a little more acetonitrile or heat results in a clear and colourless solution). This solution is added dropwise to a mixture of *n*-tetradecylphosphinic acid (2636.8 mg, 10.05 mmol, 2.1 eq.) and triethylamine (1143.5 mg, 11.3 mmol, 2.25 eq.) in dichloromethane (20 mL). The reaction mixture is heated to reflux for an hour. Afterwards, the flask is slowly cooled to room temperature and then put in a freezer. The resulting white powder is filtered on a glass frit and carefully washed by going through the slurry and break-up large chunks three times with cold methanol (40 mL). The final product is vacuum dried and yields a fine white powder (4.7 g, 93 %). Elemental analysis (CHNS) calc for C₂₈H₆₀O₄P₂Cd: %C 52.95, %H 9.54, %N 0.00, %S 0.00; found %C 51.78, %H 9.78, %N 0.49, %S 0.00.

Synthesis of cadmium selenide (CdSe) quantum dots according to Owen *et al.*²⁷ A 50 mL three neck flask, equipped with a small reflux condenser and silicon/PTFE septum, is loaded with cadmium oxide (172.1 mg, 1.34 mmol, 1.12 eq.), *n*-octadecylphosphonic acid (752.6 mg, 2.7 mmol, 2.25 eq.), and TOPO (15.0 g). The mixture is degassed (evacuated) for 30 minutes at 100 °C. The flask is filled with argon and heated to 330 °C until the CdO digests into a colourless solution. Afterwards, the temperature is lowered to 300 °C at which point 1.3 mL (1065.0 mg, 1.2 mmol, 1.00 eq.) of a premade TOP-Se solution (4.5 mmol Se per 4 gram TOP prepared stirring overnight in an argon filled glovebox) is rapidly injected. After the preferred reaction time, the mixture is cooled down to 60 °C at which point the reaction mixture is divided equally into 2 centrifuge tubes (50 mL) and the QDs are purified 3 times by precipitation with acetone (40 mL) and redispersion in toluene (5 mL) after centrifugation (10k rpm, 10 minutes).

For the synthesis with phosphinic acid or carboxylic acid, we simply replaced *n*-octadecylphosphonic acid by *n*-octadecylphosphinic acid (752.6 mg, 2.7 mmol, 2.25 eq.) or oleic acid (762.7 mg, 0.852 mL, 2.70 mmol, 2.25 eq.) respectively. Since the acidity of the phosphinic and carboxylic acid is slightly lower compared to the phosphonic acid, CdO is digested at a higher temperature (375 °C) for 4 hours until the colour of the mixture evolved into almost transparent/white suspension.

Synthesis of cadmium selenide (CdSe) nanorods according Buhro *et al.*³⁶ A 25 mL three neck flask is loaded with cadmium oxide (51.4 mg, 0.4 mmol, 1.0 eq.), di-*n*-octylphosphinic acid (203.3 mg, 0.7 mmol, 1.75 eq.), *n*-tetradecylphosphonic acid (222.7 mg, 0.8 mmol, 2.0 eq.), and TOPO (3.8g). The mixture is degassed for 40 minutes at 120 °C. The flask is filled with argon and heated to 320 °C until the CdO digests into a colourless solution. Afterwards, the temperature is lowered to 150 °C for a second degassing step for 1 hour. The flask is filled with argon and heated to 275 °C, at which point 1.2 mL of a premade TOP-Se solution (3.0 mmol Se per 5 g TOP prepared stirring overnight in an argon filled glovebox) is rapidly injected. After 30 minutes, the reaction is cooled down to 60 °C at which point the reaction mixture is transferred to a centrifuge tube (50 mL) and the NRs are purified 3 times by precipitation with acetone (40 mL) and redispersion in toluene (5mL) after centrifugation (10k rpm, 10 minutes).

For the synthesis with phosphinic acid, we replaced CdO and *n*-tetradecylphosphonic acid by presynthesized Cd(*n*-tetradecylphosphinate)₂ (254.1 mg, 0.4 mmol, 1.00 eq.). Since the phosphinic acids (*n*-tetradecyl and di-*n*-octyl) have a similar acidity they both digest the CdO resulting in a white turbid suspension.

Cadmium sulfide (CdS) NRs adapted Buhro *et al.*³⁶ Here the same recipe is followed as for the CdSe NRs but where TOP-Se is replaced by TOP-S (3.0 mmol S per 5 gram TOP).

For the synthesis with phosphinic acid, we replaced CdO and *n*-tetradecylphosphonic acid by presynthesized Cd(n-tetradecylphosphinate)₂ (254.1 mg, 0.4 mmol, 1.00 eq.) with a reaction time of 1 hour, all other parameters remained identical.

Electron Microscopy. Transmission Electron Microscopy (TEM) and High-Resolution Transmission Electron Microscopy (HRTEM) were performed on a JEOL JEM-2200FS TEM with Cs corrector operated at 200 kV (bright field).

Size and Concentration Determination. The optical band gaps of the NCs were determined by UV–vis–NIR absorption spectroscopy (PerkinElmer Lambda 950). Aliquots of approximately 0.1 mL were taken from a CdSe or CdS nanocrystal reaction and deposited into a previously weighed vial with 3 mL of hexane. Afterwards, the mixture was further diluted to obtain 10 mg mL⁻¹ reaction mixture solutions in hexane. UV-Vis absorption spectra were taken of each aliquot and the concentration of cadmium selenide in the aliquot was calculated from the size-independent absorption coefficient for CdSe at 340 nm ($\mu = 141100$ cm⁻¹) reported by Capek *et al.*⁵⁸ The size of the NCs was determined from the position of the first excitonic absorption peak using the sizing curve described by Maes *et al.*⁸⁷

NMR Spectroscopy. Nuclear Magnetic Resonance (NMR) spectra of the synthesized organics were recorded on a Bruker 300, and 400 MHz. Chemical shifts (δ) are given in ppm and the residual solvent peak was used as an internal standard (CDCl₃: δ H = 7.24 ppm, δ C = 77.06 ppm, CD₃OD: δ H = 3.31 ppm, δ C = 49.00 ppm, C₆D₆: δ H = 7.16 ppm, δ C = 128.06 ppm, toluene-d₈: δ H = 2.09 ppm, δ C = 20.43 ppm, C₆D₆: δ H = 7.16 ppm, δ C = 128.06 ppm). The signal multiplicity is denoted as follows: s (singlet), d (doublet), t (triplet), quad (quadruplet), quin (quintet), m (multiplet). Coupling constants are reported in Hertz (Hz). ¹H and ¹³C spectra were acquired using the standard pulse sequences from the Bruker library; zg30 and jmod (Attached Proton Test = APT) In the APT, the carbon resonances resulting from a -CH₂- and quaternary -C_q- are "in-phase" (orientated up), whereas the carbon resonances resulting from a

-CH₃- or -CH- are "out-of-phase" (orientated down). Although one could easily reverse the phase 180° and achieve the inverse result. ³¹P spectra of organic compounds were acquired with proton decoupling (zgpg30) and a relaxation delay (= recycle delay, or D1) of 2 seconds. All resonances were corrected prior to integration by subtracting a background from the measured intensity. The chemical shifts for other nuclei were referenced indirectly to the ¹H NMR frequency of the sample with the 'xiref'-macro in Bruker.

Nuclear Magnetic Resonance (NMR) measurements of colloidal nanocrystals were recorded on a Bruker Avance III Spectrometer operating at a ¹H frequency of 500.13 MHz and featuring a BBI probe. The sample temperature was set to 298.15. For the quantitative 1D ¹H measurements, 64k data points were sampled with the spectral width set to 16 ppm and a relaxation delay (= recycle delay, or D1) of 30 seconds to allow full relaxation of all NMR signals. The quantification was done by using the Digital ERETIC method.⁸⁸⁻⁸⁹ Onedimensional ³¹P spectra of nanocrystals were acquired using the standard pulse sequence zggpseig from the Bruker library (with a tau echo delay, *i.e.* D16 =200 μ s, and a relaxation delay (D1) of 2 seconds). DOSY measurements were performed with a double stimulated echo pulses (dstegp3s) for convection compensation and with monopolar gradient pulses,⁹⁰ and a relaxation delay (D1) of 1 second. The gradient strength was varied quadratically from 2-95% of the probe's maximum value in 64 steps, with the pulse length gradient (D20) and diffusion time (P30) optimized to ensure a final attenuation of the signal in the final increment of less than 8% relative to the first increment.

X-ray total scattering experiments. The samples were loaded in 1 mm polyimide tubes. The X-ray total scattering experiments were performed at beamline P02.1, DESY, Hamburg, Germany in rapid acquisition geometry, using a 2D detector ($150 \times 150 \mu m$ pixel size) with a sample to detector distance of 268 mm. The incident wavelength of the X-rays was $\lambda = 0.2073$ Å (59.8 keV). Calibration of the experimental setup was performed using a CeO₂ standard. Raw

2D data were corrected for geometrical effects and polarization, then azimuthally integrated to produce 1D scattering intensities versus the magnitude of the momentum transfer Q (where $Q = 4\pi \sin \theta/\lambda$ for elastic scattering) using Dioptas.⁹¹ xPDFsuite with PDFgetX3 was used to perform the background subtraction, further corrections, and normalization to obtain the reduced total scattering structure function F(Q), and Fourier transformation to obtain the pair distribution function (PDF), G(r).⁹²⁻⁹³ The following parameters were used for data reduction: $Q_{min} = 0.8 \text{ Å}^{-1}$, $Q_{max} = 17.5 \text{ Å}^{-1}$, $Q_{inst} = 17.5 \text{ Å}^{-1}$ and $R_{poly} = 0.9 \text{ Å}$. Modeling and fitting was done using Diffpy-CMI.⁹⁴

Photoluminescence. Photoluminescence spectra were measured upon excitation with a fiber coupled LED (20mA) having an emission peak wavelength of 400 nm. The samples were measured in a Spectralon coated integrating sphere. The emission spectra were analyzed with a fiber coupled Acton SP2300 monochromator and detected by a ProEM 1600 EMCCD (Princeton Instruments). The spectra were properly corrected for the spectral sensitivity of the detector part. The PLQY values were calculated by dividing the integrated number of emitted photons by the number of absorbed photons upon excitation.

Mass spectroscopy. Mass spectra (MS) are measured with an Agilent ESI single quadrupole detector type VL and an Agilent APCI single quadrupole detector type VL.

Elemental Analysis. A Thermo Scientific Flash 2000 CHNS-O analyzer equipped with a TCD detector was used to perform elemental analysis (C/H/N).

FTIR spectroscopy. Infrared spectra (IR) are recorded with a Perkin-Elmer Spectrum1000 FTIR Infrared Spectrometer.

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Associated Content

Supporting information

Provides additional HR-TEM, PDF data and results, detailed results of various experiments, ¹H NMR of surface chemistry, UV-Vis absorption spectra, PLQY spectra, and NMR spectra of synthesized compounds. This material is available free of charge via the Internet at http://pubs.acs.org/.

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Notes

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