- 1 Title: Rapid antigen test to identify COVID-19 infected patients with and without symptoms admitted
- 2 to the Emergency Department.

Introduction

- 4 Since the beginning of the SARS-CoV-2 pandemic, the rapid recognition and isolation of infected
- 5 patients have proven to be crucial factors in limiting the spread of the virus and containing the
- 6 pandemic [1, 2]. The early and accurate identification of SARS-CoV-2 patients in the emergency
- 7 department (ED) remains a major challenge [2, 3]. In addition to the confirmation or exclusion of
- 8 SARS-CoV-2 infection in symptomatic patients, the ED must simultaneously manage a large number
- 9 of patients with pathological conditions other than COVID-19 [4]. These patients may be
- asymptomatic carriers of SARS-CoV-2 infection or, if not infected, they should be separated from
- patients infected with SARS-CoV-2 [5]. The consequences of failing to correctly identify a SARS-
- 12 CoV-2-infected patient, whether symptomatic or asymptomatic, can be catastrophic [1, 5].
- 13 Currently, the diagnostic reference standard for the detection of SARS-CoV-2 is the real-time
- polymerase chain reaction (RT-PCR) test for viral RNA, primarily from orotracheal secretions [6].
- 15 Although the molecular techniques have progressed in recent months and the overall performance of
- the RT-PCR test has been improving, there are still some difficulties in its application in the daily
- 17 routine of EDs [7]. For instance, the test is not immediately available for every patient (few procedural
- sessions per day). The delay in test results and the lack of dedicated analysers in every ED are just
- 19 some of the conditions that limit the effectiveness of the RT-PCR test for COVID-19 in the
- 20 complicated clinical context of the ED [7, 8].
- 21 The US Food and Drug Administration approved the first COVID-19 antigen test, which is rapid,
- direct and economical, at the end of August 2020. This test detects specific proteins attached to the
- 23 surface of SARS-CoV-2 in samples obtained from the upper airways using an immuno-
- 24 chromatographic procedure [9]. Early laboratory evidence suggested a good correlation between
- antigen testing and RT-PCR performed on samples with a high viral load [9, 10].
- A recent systemic review synthesized the evidence on the performance of the antigen test, evaluating
- 27 943 samples from five different studies. Although the sensitivity is not optimal (average sensitivity

56.2%), the specificity above 99% may suggest a preliminary role for rapid antigen tests in a more complete clinical evaluation for the determination of SARS-CoV-2 infection [11]. The results of the studies included in the review, which were largely based on remnant laboratory samples, currently have limited clinical applicability due to the absence of evidence in real clinical contexts (e.g. the ED) and the lack of information about the symptomatic status of the patients, the timing of symptom onset and the time elapsed since possible exposure [11].

This study reports the performance of the rapid antigen test in the identification of SARS-CoV-2-infected patients in the clinical context of the ED. Both symptomatic patients suspected of having COVID-19 and asymptomatic patients who needed hospital evaluation for conditions other than

Methods

Setting and sample

COVID-19 were evaluated.

This retrospective observational study was conducted in the ED of the General Hospital of Merano (70,000 visits per year). The study period was from 1 July 2020 to 10 December 2020. This study considers patients with data previously published in the form of a preliminary report [12]. The data reported in this study are the conclusion of the previous report [12]. In July a new clinical protocol for the management of patients requiring an ED evaluation was introduced considering the use of rapid antigen tests for the timely identification patients with a SARS-CoV-2 infection. According to the clinical protocol, the rapid antigen test for SARS-CoV-2 was performed at the initial triage and at the same time as the RT-PCR swab for SARS-CoV-2 (with two different swabs) in all the patients reporting: 1) symptoms suspicious for COVID-19 infection (fever, dyspnoea, cough, sore throat, diarrhoea, vomiting, asthenia, myalgias, conjunctivitis and deficits in smell and taste); 2) all patients without COVID-19-like symptoms but with an increased temperature (>37.3°C); and 3) one positive epidemiological criterion, such as i) provenance from areas with a high incidence of SARS-

CoV-2 cases, ii) coming from other European countries (independently if as a tourist or worker), or iii) reporting contacts with a person who tested positive for SARS-CoV-2. In addition, in order to prevent the possible intra-hospital spread of the infection, we included 4) all patients evaluated in the ED for other problems not related to COVID-19 infection and who needed hospitalisation underwent both a rapid antigen test and the RT-PCR swab for SARS-CoV-2. Data from all patients who received a rapid antigen test and at least one RT-PCR swab consecutively in the ED were retrieved from the electronic database and retrospectively evaluated.

The study was conducted in accordance with the local ethical committee (Comitato etico per la sperimentazione clinica, Azienda Sanitaria dell'Alto Adige, Bolzano, Italia, approval number 57-2020) and was conducted according to the Declaration of Helsinki regarding the Ethical Principles for Medical Research Involving Human Subjects.

Identification methods and patients

All of the patients who received a rapid antigen test and at least one RT-PCR swab consecutively in the ED were retrospectively evaluated. The patient extraction procedure was performed as follows: all the electronic ED folders of patients who had performed the rapid antigenic test in the ED (computer registered request) were extracted from the ED computer database using the dedicated management software QlickView (QlikTech, Pennsylvania, PA, US). The files of the patients thus identified were manually re-evaluated by a group of ED physicians and nurses (GTu, AZ, SS, GTe, NP, DA) and only those files in which a rapid antigenic test and an RT-PCR test were registered were considered. If other RT-PCR swabs were performed after the one performed in the ED (up to a maximum of three swabs within 15 days) the results were recorded and considered. No other rapid antigen tests were performed in addition to the one performed in the ED as the test was only available in the ED for initial screening.

The rapid antigen test and the RT-PCR swab for SARS-CoV-2 were performed in the ED by appropriately trained nurses. For each patient, one nurse collected both of the swabs.

Based on the symptomatology presented at ED admission, the patients were divided into two groups: symptomatic for COVID-19 and asymptomatic for COVID-19.

The rapid antigen test for COVID-19 was performed with the STANDARD Q COVID-19 Ag (R-Ag) (SD BIOSENSOR, KR), which is a ready-to-use test, according to the manufacturer's instructions. A control line is included in the test to assess the migration of the sample. Visual interpretation of the result was performed between 15 and 30 min. The test result was reported for each patient in the personal triage record. In the case of an invalid test, a second test was performed. RT-PCR was performed with the laboratory machine GeneXpert DX System (Cepheid, CA, US), the laboratory-processed swabs were developed with XPRSARS-COV2-10 (Cepheid, CA, US). The system has been approved by the World Health Organization (WHO), and the development of RT-PCR has been conducted in accordance with the WHO guidelines. The nasopharyngeal swab is collected and placed into a transport tube containing 3 mL of saline. The specimen is briefly mixed by rapidly inverting the collection tube 5 times. Using the supplied transfer pipette, the sample is transferred to the sample chamber of the Xpert Xpress SARS-CoV-2 cartridge. The GeneXpert cartridge is loaded onto the GeneXpert DX System platform, which performs hands-off, automated sample processing, and real-time RT-PCR for detection of viral RNA.

Statistical analysis

The continuous variables are expressed as median and interquartile range (25th–75th percentile), while the categorical variables are reported as percentage and total number of events. Comparisons were made using the Mann–Whitney test, Fisher's exact test or Chi-square test, as appropriate. The performance of the antigen test was determined by analysing sensitivity, specificity and accuracy using a 2X2 table with the result of the RT-PCR test, with 95% confidence intervals (95% CI) reported.

The concordance between the antigen test and the RT-PCR test was evaluated with Cohen's kappa coefficient. The clinical benefit that may be provided by testing ED patients with antigen tests was

evaluated through decision curve analysis (DCA) [13]. DCA is a new simple statistical method that

allows calculating the clinical practicality of predictive models and can lead to clinical considerations for decision-making. DCA is a plot of net clinical benefit (y-axis) against threshold probability (x-axis) [13]. The possible net clinical benefit of performing rapid antigen tests on ED arrival is compared with the two default strategies of "all patients are infected with SARS-CoV-2" (assuming all patients are positive) and "no patient is infected with SARS-CoV-2" (assuming all patients are negative). A higher net benefit within a wide threshold range than the two standard strategies indicate the model has more potential in clinical application.

Statistical analyses were performed with STATA 13.0 software (StataCorp, College Station, TX, USA).

Results

During the study period, 3899 patients required an ED evaluation and were tested with both rapid antigen tests and RT-PCR tests (Figure 1). Of these, 30.5% (1191/3899) complained of at least one symptom possibly associated with SARS-CoV-2 infection. According to the RT-PCR results, a positive result for SARS-CoV-2 infection was found in 10.2% (397/3899) of the patients. Among the patients evaluated in the ED for a symptom that could be associated with a SARS-CoV-2 infection (n=1191), 24.7% (294/1191) tested positive for SARS-CoV-2 (86.2% in the ED, 13.8% at 48/72 hours, 0% at 14 days), and among asymptomatic patients (n=2708), 3.8% (103/2708) tested positive with the RT-PCR test (95% in the ED, 3.1% at 48/72 hours, 1.9% at 14 days). 0.2% (8/3899) reported an invalid rapid antigen test; all these patients underwent a second rapid antigen test with a valid result. The patients' demographic and clinical characteristics recorded upon ED access are reported in Table 1.

129 [Figure 1]

130 [Table 1]

The antigen test for SARS-CoV-2 was positive in 9.3% (361/3899) of all patients. Among patients with SARS-CoV-2 (n=397), rapid antigen tests were positive in 82.9% (329/397) of cases. Among

133	patients not infected by SARS-CoV-2 (n=3502), a false-positive antigen test occurred in 0.9%
134	(32/3502) of patients.
135	Overall, the sensitivity and specificity of the antigen test for detection of SARS-CoV-2 infection were
136	82.9% (95% CI, 81.0-84.8) and 99.1% (95% CI, 98.8-99.3), respectively (Table 2). The accuracy of
137	the antigen test is 97.4% (95% CI, 97.1-97.6, Cohen's $K = 0.854$, 95% CI $0.826 - 0.882$, $p < 0.001$)
138	[Table 2]
139	Performance of the SARS-CoV-2 antigen test in symptomatic and asymptomatic patients
140	The clinical characteristics of the patients evaluated in the ED for COVID-19-like symptoms are
141	reported in Table 3.
142	[Table 3]
143	A total of 3.3% (30/906) of patients with a negative result in the antigen test were found to be infected
144	with SARS-CoV-2 via the RT-PCR test, while 7.4% (21/285) of patients with positive antigen test
145	were not to be infected with SARS-CoV-2 in a RT-PCR test.
146	In symptomatic patients, the sensitivity of the antigen test for the detection of SARS-CoV-2 infection
147	was 89.8% (95% CI, 88.0-91.5), specificity was 97.6% (95% CI, 97.1-98.1) and accuracy was 95.7%
148	(95% CI, 95.1-96.3, Cohen's K = 0.884, 95% CI 0.852–0.915, p<0.001).
149	[Table 4]
150	Finally, among asymptomatic patients, the sensitivity, specificity and accuracy of the antigen test
151	compared to RT-PCR are 63.1% (95% CI, 58.4-67.8), 99.6% (95% CI, 99.5-99.7) and 98.2% (95% CI,
152	97.9-98.4, Cohen's K = 0.717, 95% CI 0.642–0.793, p<0.001).
153	
154	SARS-CoV-2 antigen test and decision curve analysis
155	In addition to the assessment of the diagnostic accuracy of the rapid antigen test, a wider evaluation on
156	the possible global clinical benefit from the use of rapid antigen test to identify COVID-19-infected

patients in ED was performed through DCA. The DCA plot seems to suggest that the use of rapid antigen tests as an initial screening tool in EDs can provide a non-negligible clinical benefit, especially when considering a population that includes asymptomatic individuals and in which the prevalence of infection appears close to its true prevalence in the general population. The inclusion of rapid antigen tests in ED had a net clinical benefit superior to clinical evaluation alone over a wide range of threshold probabilities. Between a threshold probability of 20% and 30%, the use of rapid antigen tests resulted in a net clinical benefit of between 3% and 5%, suggesting the possibility of detecting up to 5 additional true positives per 100 patients admitted in the ED who were not correctly identified by the clinical evaluation alone (Figure 2A). In fact, compared to testing all ED patients immediately with RT-PCR, at low prevalence rates (<20%) of SARS-CoV-2, the preliminary use of the rapid antigen test leads to a net clinical benefit of approximately 8%. The net clinical benefit is gradually reducing as the prevalence increases and at prevalence over 60%, indicating severe levels of infection circulating in the general population, the use of a preliminary screening test may not be a useful strategy. Moreover, clinical suspicion of SARS-CoV-2 infection based on the patients' history and signs and symptoms (clinical evaluation) does not seem to be a useful strategy for discriminating patients infected with SARS-CoV-2 admitted in ED, especially due to asymptomatic patients (Figure 2A). For prevalence values around 10%, the implementation of the antigen test strategy leads to the detection of 8 true positives per 100 RT-PCR tests performed. In addition, a hypothetical strategy involving adding the rapid antigen test to the clinical study of the symptomatology presented prior to subjecting all ED patients to the RT-PCR swab could improve by 65 out of 100 RT-PCR tests when the prevalence of SARS-CoV-2 is lower than 10% (Figure 2B).

178 [Figure 2A-B]

179

180

181

182

183

157

158

159

160

161

162

163

164

165

166

167

168

169

170

171

172

173

174

175

176

177

Discussion

Using a large cohort of patients consecutively managed in the ED, the study presents the diagnostic performance of the rapid antigen test compared to the RT-PCR swab when used in a clinical setting to identify SARS-CoV-2-infected patients in both symptomatic and asymptomatic patients managed

daily in the ED. The antigen test achieved good specificity and accuracy values, as reported in previous laboratory studies, with a fair sensitivity. The results of the present study confirm the previously published data on a partial cohort of this study, confirming the usefulness of the strategy of performing the antigen test on all patients presenting in the ED, as demonstrated by DCA [12]. Even when considering 489 more patients than in the previously published preliminary report, the performance of the rapid antigen test did not change, demonstrating a clearly superior performance in symptomatic patients compared to asymptomatic patients [12]. To the best of our knowledge, this is the study with the largest cohort that has evaluated antigen tests in daily ED clinical practice con una that has considered patients who are symptomatic as well as patients who came to the ED for other reasons and who may be asymptomatic carriers of the virus. Since the hospitalisation of one of these patients (asymptomatic carriers) incorrectly identified as SARS-CoV-2 negative can lead to dramatic consequences, the evaluation of appropriate strategies in order to prevent this is crucial [14, 15]. Some of the characteristics of the rapid antigen test described at the time of commercialisation seem to adapt well to the ED scenario [9, 11]. Its simplicity, repeatability, rapidity of execution and, more importantly, immediate results could overcome the limitations presented by the RT-PCR swab, adapting better to the dynamic nature of EDs [9, 10, 11]. The first laboratory studies for the validation of the methodology have provided good initial indications for different types of antigen tests. The laboratory comparison with RT-PCR performed on the same microbiological sample revealed a sensitivity of 68%, a specificity of 100% and an accuracy of 72% for the antigen test [16]. Only 31 out of 239 patients were negative in the detection of the viral RNA, indicating a high prevalence of the disease in the limited study cohort [16]. In addition to the high prevalence of infection, the absence of clinical information may limit the application of these results in clinical practice. Similarly, Porte et al., who tested 127 samples and found 82 to be positive by RT-PCR, confirmed a 100% specificity of the antigen test and indicated a sensitivity and accuracy of 93.9% and 96.1%, respectively [10]. Despite the high prevalence of SARS-CoV-2 in the present study, the sensitivity of the antigen test seems to improve as the viral load presented in the samples

184

185

186

187

188

189

190

191

192

193

194

195

196

197

198

199

200

201

202

203

204

205

206

207

208

209

increases. In the case of reduced prevalence and reduced viral load, the antigen test seems to have a decreased ability to identify patients infected with SARS-CoV-2 [10, 17]. More recently, Cerutti et al. reported the first indications of the antigen test in a group of asymptomatic patients (travellers returning from risk areas) [18]. The sensitivity and specificity of the antigen test in this group of asymptomatic patients were 40% and 100%, respectively, with a RT-PCR swab agreement close to 98%, higher than in the symptomatic group [18]. An initial systematic review of the performance of the antigen test was conducted in an attempt to provide precise indications that can be applied in clinical practice [11]. By grouping 943 tests from five studies, the sensitivity of the antigen tests was found to be relatively low (56.2%, 95% CI 29.5%-79.8%), but a consistently high level of specificity was observed (mean 99.5%, 95% CI 98.1%–99.9%) [11]. However, the absence of information on patient symptomatology limits the evaluation of the usefulness of the antigen test in clinical practice. The findings of the current study are the first to translate the evidence from previous laboratory studies into clinical practice. Due to the fact that the consequences of a single error in the detection of a SARS-CoV-2-infected patient accessing the hospital can have catastrophic consequences, in daily ED practice, the confirmed high levels of specificity do not seem to be sufficient to compensate for the sub-optimal levels of sensitivity and to provide perfect flow management based only on antigen tests [14, 15, 19]. However, at the indicated levels of prevalence, a positive antigen test in symptomatic patients could allow early and rapid identification of the infection while, in case of a negative antigen test, the maintenance of a high clinical suspicion could guarantee safe management until successive microbiological confirmations. Moreover, in asymptomatic patients, the use of the antigen test, which alone cannot guarantee that all SARS-CoV-2-positive patients are identified, could allow an immediate identification of a portion of patients and thus improve the entire ED decision-making process. The ability of asymptomatic patients to spread the infection is well known [14, 20, 21]. Hospitalisation of an undiagnosed asymptomatic patient in a non-COVID-19 department can lead to intra-hospital spread of the virus, affecting patients and healthcare workers [14, 20, 21]. The development of specific strategies for these patients is important. The use of the antigen test in the ED, as suggested by the DCA, provides a net clinical benefit in the process of identifying infected

211

212

213

214

215

216

217

218

219

220

221

222

223

224

225

226

227

228

229

230

231

232

233

234

235

236

237

individuals. Although it is still far from the performance of an 'ideal test', the antigen test seems to be a good initial strategy in both symptomatic patients, in whom it is combined with clinical evaluation, and asymptomatic patients, where it can immediately identify a portion of patients who otherwise would not have been rapidly identified.

The study presents some limitations. First, there are no quantitative data available about the viral load to confirm a possible condition of reduced viral load in the false-negative antigen tests. Second, there

to confirm a possible condition of reduced viral load in the false-negative antigen tests. Second, there are no analyses available about the risk of viral transmission by patients who received a false-negative antigen test. Third, asymptomatic patients who were discharged after ED evaluation and did not require hospitalisation were not included in the study because they were not subjected to a RT-PCR swab.

Conclusions

Rapid and accurate identification of patients with SARS-CoV-2 infection in the ED is essential to contain the progression of the pandemic and keep the hospital secure. The overall sensitivity, specificity and accuracy of the rapid antigen test were very good, yet with low sensitivity in asymptomatic patients. The assessment of patients admitted to the ED for a SARS-CoV-2 infection during the initial triage via rapid antigen test has an additional clinical benefit. In symptomatic patients arriving in the ED, where the prevalence of SARS-CoV-2 infection is higher, a positive antigen test can accelerate and optimise the management of the infected patient. A negative antigen test in these symptomatic patients should however be followed by RT-PCR testing to confirm the absence of disease. In asymptomatic patients, for whom more secure and precise strategies need to be implemented, an initial antigen test seems to identify patients with SARS-CoV-2 infection who otherwise would not have been rapidly detected. However, a negative rapid antigen test in asymptomatic patients is not sufficient to safely exclude SARS-CoV-2 infection.

287

284

285

286

10.3390/ijerph17228302.

Season, COVID-19 Epidemic, and Lockdown Periods in View of Managing a Future Disaster Risk: A

Multicenter Observational Study. Int J Environ Res Public Health. 2020 Nov 10;17(22):E8302. doi:

- 288 5) Yu X, Yang R. COVID-19 transmission through asymptomatic carriers is a challenge to
- 289 containment. Influenza Other Respir Viruses. 2020 Jul;14(4):474-475. doi: 10.1111/irv.12743. Epub
- 290 2020 Apr 15.

291

- 292 6) Wu S.Y, Yu M.Y, Tsang H.F, Chan L.W.C, Cho W.C.S, Yu A.K.Y, Li M.J.W, Wong Y.K.E, Pei
- 293 X.M, Wong S.C.C. The diagnostic methods in the COVID-19 pandemic, today and in the future.
- 294 Expert Rev Mol Diagn. 2020 Sep;20(9):985-993. doi: 10.1080/14737159.2020.1816171. Epub 2020
- 295 Sep 16.

296

- 297 7) Zitek T. The Appropriate Use of Testing for COVID-19. West J Emerg Med. 2020 Apr
- 298 13;21(3):470-472. doi: 10.5811/westjem.2020.4.47370.

299

- 300 8) Carpenter C.R, Mudd P.A, West C.P, Wilber E, Wilber S.T. Diagnosing COVID-19 in the
- 301 Emergency Department: A Scoping Review of Clinical Examinations, Laboratory Tests, Imaging
- Accuracy, and Biases. Acad Emerg Med. 2020 Jun 16;10.1111/acem.14048. doi:
- 303 10.1111/acem.14048.

304

- 9) Weissleder R, Lee H, Ko J, Pittet M.J. COVID-19 diagnostics in context. Sci Transl Med. 2020 Jun
- 306 3;12(546):eabc1931. doi: 10.1126/scitranslmed.abc1931.

307

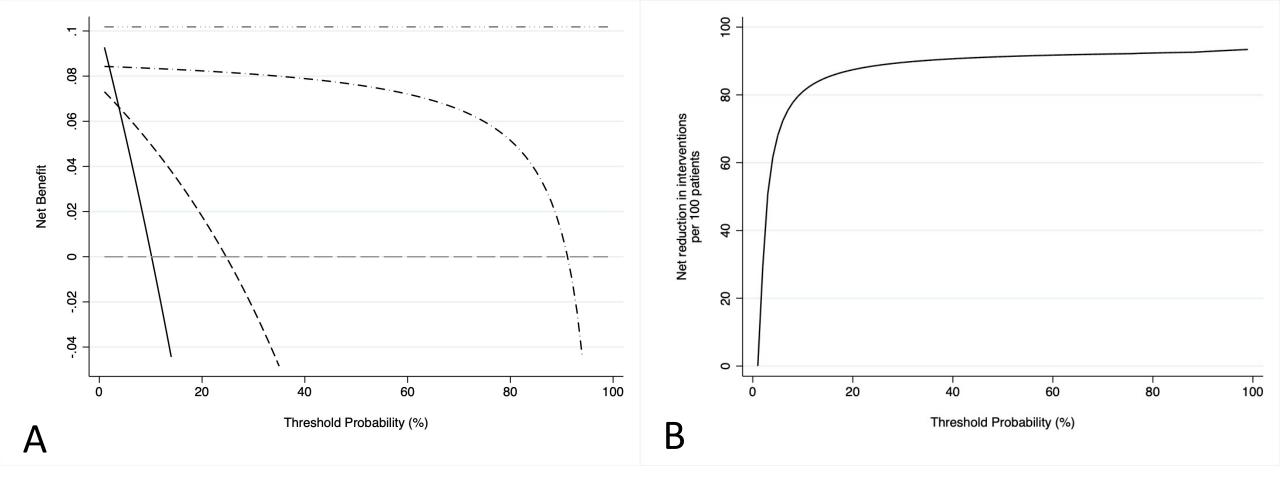
- 308 10) Porte L, Legarraga P, Vollrath V, Aguilera X, Munita J.M, Araos R, Pizarro G, Pablo V,
- 309 Iruretagoyena M, Dittrich S, Weitzel T. Evaluation of a novel antigen-based rapid detection test for the
- diagnosis of SARS-CoV-2 in respiratory samples. Int J Infect Dis. 2020 Oct;99:328-333. doi:
- 311 10.1016/j.ijid.2020.05.098. Epub 2020 Jun 1.

11) Dinnes J, Deeks J.J, Adriano A, Berhane S, Davenport C, Dittrich S, Emperador D, Takwoingi Y, 313 Cunningham J, Beese S, Dretzke J, Ferrante di Ruffano L, Harris I.M, Price M.J, Taylor-Philips S, 314 315 Hooft L, Leeflang M.M, Spijker R, Van den Bruel A. Rapid, point-of-care antigen and molecularbased tests for diagnosis of SARS-CoV-2 infection. Cochrane Database Syst Rev. 2020 Aug 316 317 26;8:CD013705. doi: 10.1002/14651858.CD013705. 318 319 12) Turcato G, Zaboli A, Pfeifer N, Ciccariello L, Sibilio S, Tezza G, Ausserhofer D. Clinical 320 application of a rapid antigen test for the detection of SARS-CoV-2 infection in symptomatic and asymptomatic patients evaluated in the emergency department: A preliminary report. J Infect. 2021 321 322 Mar;82(3):e14-e16. doi: 10.1016/j.jinf.2020.12.012. 323 324 13) Vickers A.J, van Calster B, Steyerberg E.W. A simple, step-by-step guide to interpreting decision 325 curve analysis. Diagn Progn Res. 2019 Oct 4;3:18. doi: 10.1186/s41512-019-0064-7. eCollection 326 2019. 327 14) Lavezzo E, Franchin E, Ciavarella C, Cuomo-Dannenburg G, Barzon L, Del Vecchio C, Rossi L, 328 Manganelli R, Loregian A, Navarin N, Abate D, Sciro M, Merigliano S, De Canale E, Vanuzzo M.C, 329 Besutti V, Saluzzo F, Onelia F, Pacenti M, Parisi S.G, Carretta G, Donato D, Flor L, Cocchio S, Masi 330 331 G, Sperduti A, Cattarino L, Salvador R, Nicoletti M, Caldart F, Castelli G, Nieddu E, Labella B, Fava 332 L, Drigo M, Gaythrope K.A.M, Brazzale A.R, Toppo S, Trevisan M, Baldo V, Donnelly C.A, Ferguson N.M, Dorigatti I, Crisanti A. Suppression of a SARS-CoV-2 outbreak in the Italian 333 municipality of Vo'. Nature. 2020 Aug;584(7821):425-429. doi: 10.1038/s41586-020-2488-1. Epub 334 335 2020 Jun 30.

15) Al-Sadeq D.W, Nasrallah G.K. The incidence of the novel coronavirus SARS-CoV-2 among 337 asymptomatic patients: A systematic review. Int J Infect Dis. 2020 Sep;98:372-380. doi: 338 339 10.1016/j.ijid.2020.06.098. Epub 2020 Jul 2. 340 16) Diao B, Wen K, Chen J, Liu Y, Yuan Z, Han C, Chen J, Pan Y, Chen L, Dan Y, Wang J, Chen Y, 341 Deng G, Zhou H, Wu Y. Diagnosis of Acute Respiratory Syndrome Coronavirus 2 Infection by 342 343 Detection of Nucleocapsid Protein. doi: https://doi.org/10.1101/2020.03.07.20032524. 344 345 17) Hirotsu Y, Maejima M, Shibusawa M, Nagakubo Y, Hosaka K, Amemiya K, Sueki H, Hayakawa 346 M, Mochizuki H, Tsutsui T, Kakizaki Y, Miyashita Y, Yagi S, Kojima S, Omata M. Comparison of 347 automated SARS-CoV-2 antigen test for COVID-19 infection with quantitative RT-PCR using 313 348 nasopharyngeal swabs, including from seven serially followed patients. Int J Infect Dis. 2020 Oct; 99: 349 397–402. Published online 2020 Aug 12. doi: 10.1016/j.ijid.2020.08.029 350 351 18) Cerutti F, Burdino E, Milia M.G, Allice T, Gregori G, Bruzzone B, Ghisetti V. Urgent need of rapid tests for SARS CoV-2 antigen detection: Evaluation of the SD-Biosensor antigen test for SARS-352 CoV-2. J Clin Virol. 2020 Nov;132:104654. doi: 10.1016/j.jcv.2020.104654. Epub 2020 Sep 29. 353 354 19) Scohy A, Anantharajah A, Bodéus M, Kabmba-Mukadi B, Verroken A, Rodriguez-Villalobos H. 355 356 Low performance of rapid antigen detection test as frontline testing for COVID-19 diagnosis. J Clin Virol. 2020 Aug;129:104455. doi: 10.1016/j.jcv.2020.104455. Epub 2020 May 21. 357 358 359 20) Gao M, Yang L, Chen X, Deng Y, Yang S, Xu H, Chen Z, Gao X. A study on infectivity of 360 asymptomatic SARS-CoV-2 carriers. Respir Med. 2020 Aug;169:106026. doi: 10.1016/j.rmed.2020.106026. Epub 2020 May 13. 361

21) Huang L, Zhang X, Zhang X, Wei Z, Zhang L, Xu J, Liang P, Xu Y, Zhang C, Xu A. Rapid asymptomatic transmission of COVID-19 during the incubation period demonstrating strong infectivity in a cluster of youngsters aged 16-23 years outside Wuhan and characteristics of young patients with COVID-19: A prospective contact-tracing study. J Infect. 2020 Jun;80(6):e1-e13. doi: 10.1016/j.jinf.2020.03.006. Epub 2020 Apr 10.

Figure 2: A) Decision curve analysis and its distribution. Grey dashed line: assume no patients have COVID-19. Black line: assume all patients have COVID-19. Grey dash-dotted line: a hypothetical perfect test. Black dashed line: the strategy of discovering COVID-19-infected patients only on the basis of their symptoms. Black dash-dotted line: the strategy of performing antigen tests on patients in the ED. The X-axis indicates the threshold probability for cardiological adverse event and the Y-axis indicates the net benefit. The black line assumes that all the patients would be SARS-CoV-2 infected, while the grey line reflects the assumption that no patients would be SARS-CoV-2 infected. The dashed black line represents the net clinical benefit provided by the clinical evaluation and the grey dashed and dotted line represents the net clinical benefit provided by the introduction of rapid antigen tests in the ED. As demonstrated in the graph, rapid antigen tests achieved greater clinical utility in the threshold probability, indicating that rapid antigen tests may be a valuable tool in identifying SARS-CoV-2 positive patients. B) Decision curve analysis plotting the decrease in RT-PCR swabs due to the clinical evaluation plus the implementation of the antigen test based on the prevalence of SARS-CoV-2 in the population.



Variable	Global	COVID-19 negative	COVID-19 positive	p
Patients, n (%)	3899 (100)	3502 (89.8)	397 (10.2)	
Sex, n (%)				0.501
Male	1992 (51.1)	1789 (51.1)	212 (53.4)	
Female	1907 (48.9)	1713 (48.9)	185 (46.6)	
Age, years, median (IQR)	69 (49-82)	69 (49-82)	68 (47-81)	0.830
Paediatrics population*, n (%)	91 (2.3)	85 (2.4)	6 (1.5)	0.296
Elderly population ^{\$} , n (%)	1866 (47.9)	1672 (47.7)	194 (48.9)	0.672
Arrival mode, n (%)				0.005
Walk-in/Private vehicle	1740 (44.6)	1612 (46.0)	128 (32.2)	
Ambulance	1798 (46.1)	1551 (44.3)	247 (62.2)	
Emergency medical service	361 (9.3)	339 (9.7)	22 (5.5)	
Days of the week, n (%)				0.132
During the week	2998 (76.9)	2705 (77.2)	293 (73.8)	
Weekend	901 (23.1)	797 (22.8)	104 (26.2)	
Access during night (20.00-08.00), n (%)	789 (20.2)	718 (20.5)	71 (17.9)	0.236
Tourist, n (%)	235 (6.0)	227 (6.0)	8 (2.0)	< 0.001
Triage code, n (%)				0.016
Blue and Green	1666 (42.7)	1469 (41.9)	197 (49.6)	
Yellow	1501 (38.5)	1369 (39.1)	132 (33.2)	
Orange and Red	732 (18.8)	664 (19.0)	68 (17.1)	
COVID-19 symptoms	1191 (30.5)	897 (25.6)	294 (74.1)	< 0.001
Area of treatment, n (%)				< 0.001
Surgical area	1032 (26.5)	985 (28.1)	47 (11.8)	
Internal medicine area	2279 (58.5)	1945 (55.5)	334 (84.1)	
Gynaecological area	189 (4.8)	182 (5.2)	7 (1.8)	
Trauma area	399 (10.2)	390 (11.1)	9 (2.3)	
Rapid antigen test for COVID-19, n (%)				< 0.001
Negative antigen test	3538 (90.7)	3470 (99.1)	68 (17.1)	
Positive antigen test	361 (9.3)	32 (0.9)	329 (82.9)	

Table 1: Demographic and baseline characteristics of all enrolled patients, divided according to RT-PCR result for SARS-CoV-2. * =less than 14 years; \$=over than 65 years.

non-infected	infected	
3,470	68	
32	329	
82.9% (81.0-84.8)		
99.1% (98.8-99.3)		
91.1% (89.7-92.5)		
98.1% (97.8-98.3)		
97.4% (97.1-97.6)		
	32 82.9% (8 99.1% (9 91.1% (8 98.1% (9	

Table 2: One 2×2 contingency table on rapid antigen test performance in the assessment of patients with COVID-19 among all patients.

Variable	Negative antigen test	Positive antigen test	p
	for COVID-19	for COVID-19	
Patients, n (%)	906 (76.1)	285 (23.9)	
SARS-CoV-2 positive	30 (3.3)	264 (92.6)	< 0.001
Age, years, median (IQR)	75 (58-84)	68 (50-8)	0.002
Sex, n (%)			0.556
Male	486 (53.7)	161 (56.4)	
Female	420 (46.3)	124 (43.6)	
COVID-19 symptoms, n (%)			
Fever or history of fever	451 (52.7)	148 (53.2)	0.890
Cough	112 (13.2)	84 (30.2)	< 0.001
Dyspnoea	327 (38.2)	114 (41.2)	0.395
Gastroenterological	204 (23.8)	61 (21.9)	0.568
Other symptoms	369 (43.2)	137 (49.5)	0.071
Time of onset of symptoms, days, median (IQR)	2 (1-3)	2 (2-4)	< 0.001
Comorbidity, n (%)	415 (48.5)	91 (33.0)	< 0.001

Table 3: Demographic and baseline characteristics of the patients evaluated in the ED for symptoms suspicious for SARS-CoV-2.

Only considering symptomatic patients for COVID-19					
	Patients COVID-19	Patients COVID-19			
	non-infected	infected			
Negative antigen test for COVID-19	876	30			
Positive antigen test for COVID-19	21	264			
Sensitivity	89.8% (88.0-91.5)				
Specificity	97.6% (97.1-98.1)				
Positive predictive value	92.6% (91.0-94.1)				
Negative predictive value	96.7% (96.0-97.2)				
Accuracy (correctly classified)	95.7% (95.1-96.3)				
Only considering asymptomatic patients for COVID-19					
	Patients COVID-19	Patients COVID-19			
	non-infected	infected			
Negative antigen test for COVID-19	2,594	38			
Positive antigen test for COVID-19	11	65			
Sensitivity	63.1% (58.4-67.8)				
Specificity	99.6% (99.5-99.7)				
Positive predictive value	85.5% (81.5-89.5)				
Negative predictive value	98.5% (98.3-98.7)				
Accuracy (correctly classified)	98.2% (97.9-98.4)				

Table 4: Two 2 × 2 contingency tables on antigen rapid test performance in the assessment of patients with COVID-19. The first table focuses on the rapid antigen test among symptomatic patients for SARS-CoV-2 and the table below focuses on the rapid antigen test among asymptomatic patients for SARS-CoV-2.