

SHORT REPORT

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In vitro and *in vivo* antischistosomal activity of ferroquine derivatives

Jennifer Keiser^{1,2*}, Mireille Vargas^{1,2}, Riccardo Rubbiani³, Gilles Gasser³ and Christophe Biot^{4,5}

Abstract

Background: Schistosomiasis is a neglected tropical disease and drug-repurposing is a useful strategy to fill its exhausted drug development pipeline. The ferrocenyl analogue of chloroquine, ferroquine, is an antimalarial in late-stage drug development. The aim of the present work was to study the antischistosomal activity of ferroquine against *Schistosoma mansoni* adult worms and newly transformed schistosomula (NTS) *in vitro* and *in vivo*. Hydroxyl-ferroquine and ruthenoquine were included to study the potential role of reactive oxygen species in the antischistosomal activity. Chloroquine and mefloquine, the later described for its antischistosomal properties, served as comparators.

Findings: All metal complexes were shown to be moderately cytotoxic on human cervix HeLa cancer cells and human fetal lung fibroblasts MRC-5. 72 hours post-incubation NTS exposed to 33.3 μ M ruthenoquine had died, while ferroquine and hydroxyl-ferroquine treated worms were strongly affected. No activity was observed treating NTS with chloroquine at 33.3 μ M. Incubation of adult *S. mansoni* with 33.3 μ M of the organometallic derivatives were highly affected in viability but were still alive 72 hours post-incubation. Mefloquine showed the highest activity against NTS and adult *S. mansoni*. Low total worm burden reductions of 0-36% were observed following oral administration of 200-800 mg/kg of the ferroquine derivatives to *S. mansoni*-infected mice.

Conclusions: The organometallic compounds evaluated in this study revealed moderate *in vitro* activity against both larval and adult stages of *S. mansoni* but low *in vivo* activity. No correlation can be drawn between the antimalarial and antischistosomal activity of chloroquine analogues and oxidative shock does not seem to play a role in the activity of these compounds against *S. mansoni*.

Keywords: *Schistosoma mansoni*, Ferroquine, Chloroquine, Ruthenoquine, Hydroxyl-ferroquine, *In vitro*, *In vivo*

Background

Schistosomiasis is a neglected tropical disease, which causes a considerable public health burden [1]. Millions of people require treatment each year to prevent the considerable health effects of schistosomiasis. For example, in 2012 it has been estimated that 249 million people should be treated in preventive chemotherapy programs [2]. Yet, treatment and control of schistosomiasis relies on a single drug, namely praziquantel [1,3,4].

Research on novel antischistosomal drugs is nearly exclusively driven by academic groups with a strong focus on phenotypic approaches, since protein targets

are almost unknown [5]. Over the past years, libraries consisting of existing drugs, so called drug-repurposing, were investigated in order to fill the exhausted drug development pipeline for schistosomiasis [6]. Drug repurposing circumvents the high cost of drug discovery and development, the high failure rates and the long duration to develop novel treatments by finding new uses for compounds other than those they were initially intended to treat [7]. Antimalarials, in particular, have been well studied, which is not surprising since both *Plasmodium* and schistosomes degrade hemoglobin [4]. For example, the excellent antischistosomal activity of mefloquine (MQ, Figure 1) was demonstrated in *S. haematobium*, *S. japonicum* and *S. mansoni* rodent models [8]. The promising results obtained with MQ in laboratory studies were followed up in clinical trials in the past years [9,10]. Over the last decade, there has

* Correspondence: jennifer.keiser@unibas.ch

¹Department of Medical Parasitology and Infection Biology, Swiss Tropical and Public Health Institute, Basel, Switzerland

²University of Basel, Basel, Switzerland

Full list of author information is available at the end of the article

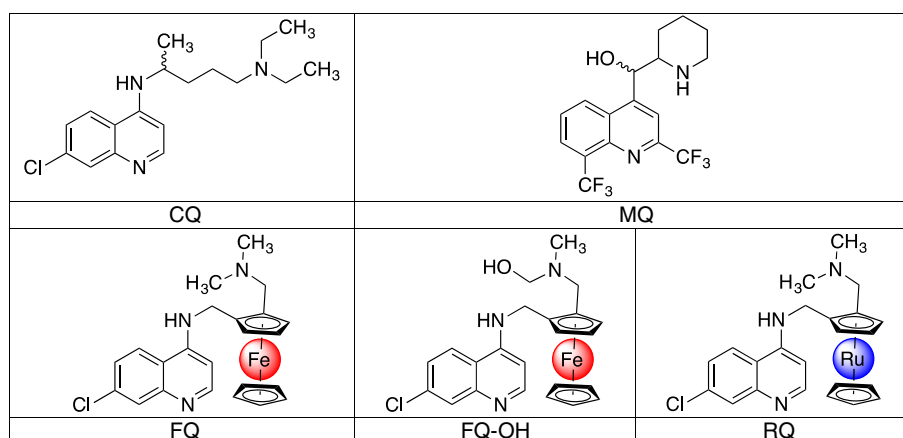


Figure 1 Chemical structures of chloroquine (CQ), mefloquine (MQ), ferroquine (FQ), hydroxyl-ferroquine (FQ-OH) and ruthenoquine (RQ).

been increasing interest in metal-based drugs and numerous metal complexes have been synthesized and evaluated as antimalarial agents [11]. The most promising candidate was undoubtedly the ferrocenyl analogue of chloroquine (CQ, Figure 1), ferroquine (FQ, Figure 1). FQ reached two different Phase IIb clinical trials, in combination with artesunate or OZ439, for the treatment of uncomplicated *Plasmodium falciparum* malaria (<http://www.mmv.org/research-development/project-portfolio/oz439fq>). FQ has a specific parasitocidal effect on *Plasmodium* due to the FQ-induced oxidative stress within the digestive vacuole and the subsequent destruction of the membrane resulting in death of the parasite [12].

The aim of the present work was to study the antischistosomal activities of FQ against *S. mansoni* *in vitro* and *in vivo*. In addition, we investigated whether redox activation plays a role in antischistosomal activity by including the ruthenocenyl analogue of FQ, namely ruthenoquine (RQ, Figure 1), which was shown, contrary to FQ, to be unable to produce reactive oxygen species (ROS) in *P. falciparum* [13]. This difference in redox activity was assumed to be responsible for the greater antimalarial activity of FQ over RQ. Furthermore, the antischistosomal activity of hydroxyl-ferroquine (FQ-OH, Figure 1) was also studied since FQ-OH is also able to produce hydroxyl radicals and provides reduced cytotoxic effects compared to FQ [14].

Findings

Methods

Animals and parasites

In vivo studies were carried out in accordance with Swiss national and cantonal regulations on animal welfare (permission no. 2070) at the Swiss Tropical and Public Health Institute (Basel, Switzerland). Female mice (NMRI strain, $n = 31$; weight ~ 20 – 22 g) were purchased

from Charles River, Germany, kept under environmentally-controlled conditions (temperature $\sim 25^{\circ}\text{C}$; humidity $\sim 70\%$; 12-hour light and 12-hour dark cycle) with free access to water and rodent diet and acclimatized for one week before infection. Cercariae of *S. mansoni* were obtained from infected intermediate host snails (*Biomphalaria glabrata*) as described previously [15].

Compounds

FQ, FQ-OH and RQ were synthesized according to reported procedures [13,14,16]. MQ was kindly obtained from Mepha AG (Aesch, Switzerland). CQ was purchased from Sigma (Buchs, Switzerland). For *in vitro* studies, compounds were dissolved in DMSO (Fluka, Buchs, Switzerland) to obtain stock solutions of 10 mg/ml. For *in vivo* studies, compounds were suspended in 7% (v/v) Tween 80 and 3% (v/v) ethanol shortly before oral treatment (10 ml/kg) of mice.

In vitro studies

Newly transformed schistosomula (NTS)

S. mansoni cercariae were mechanically transformed to newly transformed schistosomula (NTS) [17]. A NTS suspension at a concentration of 100 NTS per 50 μl was prepared using Medium 199 (Invitrogen, Carlsbad, CA) [supplemented with 5% inactivated fetal calf serum (iFCS) and 100 U/ml penicillin and 100 mg/mL streptomycin (Invitrogen)]. NTS were incubated with 10 μM and 33.3 μM of the test compounds for 72 h. Compounds were tested at least in triplicate and the highest concentration of DMSO served as control. Plates were incubated at 37°C , 5% CO_2 . NTS were evaluated by microscopic readout (Carl Zeiss, Germany, magnification 80x) using a viability scale scoring death, changes in motility, viability, and morphological alterations [17].

Adult *S. mansoni*

Adult schistosomes obtained from infected mice were incubated in the presence of 10 μ M and 33.3 μ M of the test compounds for up to 72 h. Phenotypes were monitored daily based on motility, viability and morphological alterations under an inverse microscope (Carl Zeiss, Germany, magnification 80 \times).

Cytotoxicity studies

Cytotoxicity studies were performed on human cervix HeLa cancer cells and non tumorigenic human fetal lung fibroblasts MRC-5 to compare the activity of FQ, RQ, FQ-OH, CQ, MQ and cisplatin. The cell viability was determined via a colorimetric cell-based assay using Resazurin (Promocell GmbH). Briefly, one day before treatment cells were plated in triplicates in 96-well plates at a density of 4×10^3 cells/well in 100 μ l. Upon treating cells with increasing concentrations of the target complexes (freshly prepared stock solution in DMSO), cells were incubated at 37°C/6% CO₂ for 48 h, the medium was removed, and 100 μ l of complete medium containing resazurin (0.2 mg/ml final concentration) was added. After 4 h of incubation at 37°C/6% CO₂, the fluorescence of the highly red fluorescent resorufin product was quantified at 590 nm emission with 540 nm excitation wavelength in a SpectraMax M5 microplate Reader.

S. mansoni in vivo studies

Groups of 3–4 NMRI mice were treated orally with single oral doses of 200 mg/kg of FQ, FQ-OH and RQ. In addition, one group of mice was treated with a single oral dose of 800 mg/kg FQ. Untreated mice served as controls in all experiments. At 21 d post-treatment, animals were killed by the CO₂ method and dissected. Worms were removed by picking, then sexed and counted as previously described [18].

Statistics

Parasite viability values of treated and untreated worms obtained from microscopic evaluation were averaged (means (+/- standard deviation)) using Microsoft Excel software. The Kruskal-Wallis test was applied for *in vivo* studies, comparing the medians of the worm counts of the treatment and control groups. A difference in median was considered to be significant at a significance level of 5% (StatsDirect statistical software, version 2.7.2.; StatsDirect Ltd., United Kingdom).

Results and discussion

Cytotoxicity studies

We assessed the potential toxicity of the compounds studied in this work on cervical cancer cells (HeLa) and non-cancerous cells (MRC-5). Cisplatin, the best known metal-based drug on the market was used as a reference.

As shown in Table 1, all metal complexes were shown to be moderately cytotoxic. MQ was the most toxic compound in these assays. This is not surprising, since MQ is well known for its adverse events, including gastrointestinal effects [9] and neuropsychiatric toxicity [19]. On the other hand, FQ was well tolerated and shown to be lacking relevant adverse effects on central nervous system, respiratory, renal, and gastrointestinal functions in a phase I trial [20]. However, larger trials are necessary to confirm this finding.

Activity against NTS and adult *S. mansoni* in vitro

24 hours post-incubation all NTS exposed to 33.3 μ M FQ, FQ-OH and RQ showed strongly reduced viabilities. 72 hours post-incubation all NTS exposed to 33.3 μ M RQ had died, while FQ and FQ-OH treated worms were strongly affected but still alive. Therefore, unlike for *P. falciparum* and *Trypanosoma brucei gambiense* [21], oxidative shock does not seem to play a role in the activity of these compounds against *S. mansoni*, since RQ, which cannot produce ROS is the most active of the three organometallic drug against NTS *in vitro*. This absence of redox activation could be explained by the difference in the target of FQ, FQ-OH and RQ in *S. mansoni* compared to *P. falciparum*. In *P. falciparum*, sour conditions present in the acidic compartment where FQ localizes as well as of H₂O₂ are necessary for the formation of HO \cdot [12]. For comparison, no activity was observed treating NTS with CQ at 33.3 μ M (Figure 2a) while at this concentration 24 hours post-incubation with MQ all worms had died. The antischistosomal mechanism of action of MQ and CQ is also not known. Note that these 2 drugs do not produce hydroxyl radicals [22]. The antimalarial activity of CQ and MQ is largely attributed to an inhibition of hemoglobin degradation. However, it was recently shown that hemozoin inhibition of CQ, MQ or quinine does not exhibit a correlation with their antischistosomal properties [23]. Furthermore, it was demonstrated that MQ interferes with glycolysis in NTS [24].

Table 1 IC₅₀ values for ferroquine (FQ), hydroxyl-ferroquine (FQ-OH), ruthenoquine (RQ), chloroquine (CQ) and mefloquine (MQ) in non-cancerous MRC-5 and HeLa cancer cells

Complex	IC ₅₀ MRC-5 (μ M)	IC ₅₀ HeLa (μ M)
Cisplatin	7.9 \pm 1.2	11.5 \pm 2.9
FQ	24.4 \pm 0.9	10.1 \pm 0.3
FQ-OH	22.6 \pm 1.2	16.8 \pm 1.5
RQ	21.9 \pm 2.6	8.8 \pm 0.41
CQ	55.1 \pm 1.3	87.0 \pm 1.5
MQ	16.7 \pm 0.2	6.7 \pm 0.7

Cisplatin was used as positive control. Standard deviations are shown in superscript.

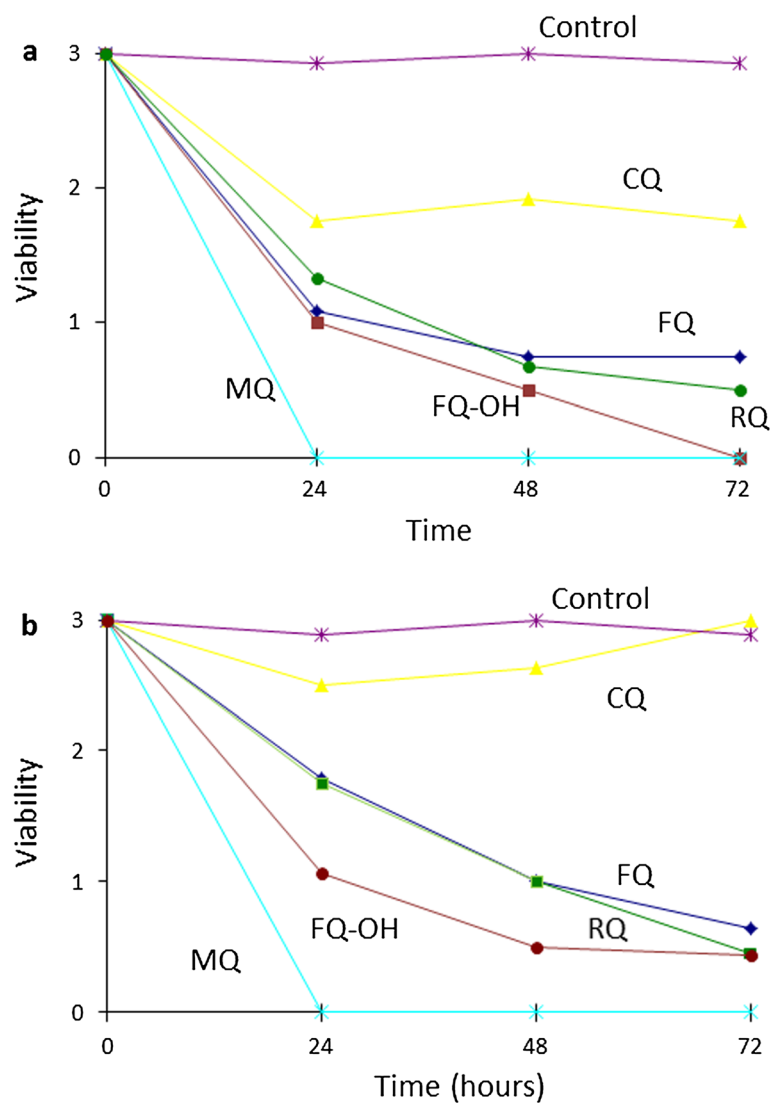


Figure 2 Activity of ferroquine (FQ), hydroxy-ferroquine (FQ-OH), ruthenoquine (RQ), chloroquine (CQ) and mefloquine (MQ) versus untreated control worms in vitro at 33.3 µg/ml against NTS (a) and adult *S. mansoni* (b).

Exposure to 10 µM of the test drugs resulted in reduced motilities of NTS but the larvae did not die (data not shown).

A similar trend was observed on adult *S. mansoni*. Incubation of adult *S. mansoni* with 33.3 µM of MQ resulted in death of worms 24 h post-treatment, while adult *S. mansoni* exposed to the organometallic derivatives were highly affected in viability but were still alive 72 hours post-incubation (Figure 2b). At 10 µM adults showed strongly reduced viabilities 48–72 hours post-incubation with all derivatives (data not shown).

Activity against adult *S. mansoni* in vivo

Compounds progressed into *in vivo* studies despite the fact that *in vitro* activities were only observed at concentrations that were close to cytotoxic concentrations.

However, in our recent studies MQ was well tolerated by mice [8].

We have previously shown that CQ lacked antischistosomal activity *in vivo* (worm burden reduction of 11%) [8]. FQ is known for important and additional properties (e.g. its higher lipophilicity) and mechanism of actions (the above mentioned radical formation as well as hemozoin inhibition) compared to CQ [22]. Yet, treatment of mice with 200 and 800 mg/kg FQ, showed low total worm burden reductions of 19.4% and 35.6% (Table 2). One of the mice treated with 800 mg/kg FQ died within 24 hours post-treatment. No activity was observed treating mice with RQ at 200 mg/kg. Finally, a total worm burden reduction of 17.3% was observed following treatment with FQ-OH. Hence, modification of CQ by a ferrocenyl or ruthenocenyl fragment does not increase the

Table 2 Effect on worm burden of single oral doses of three selected organometallic CQ derivatives administered to mice harboring a 49-day-old adult *S. mansoni* infection, stratified by sex and worm distribution

Drug	Dose (mg/kg)	No. of mice investigated	No. of mice cured	Mean number of worms (SD)					Total worm burden reduction (%)	p-value	Female worm burden reduction (%)	p-value
				Liver	Mesenteric veins	Total	Males	Females				
Control ¹	-	8	-	0.4 (0.7)	33.8 (10.2)	34.1 (10.3)	19.9 (7.7)	14.3 (4.2)	-	-	-	-
Control ²	-	8	-	0.6 (1.2)	25.8 (16.7)	26.4 (16.7)	14.5 (9.3)	11.9 (7.7)	-	-	-	-
FQ ^a	200	4	0	1.8 (2.4)	25.8 (7.3)	27.5 (7.3)	16.3 (4.9)	11.3 (4.0)	19.4	>0.05	21.0	>0.05
FQ ^b	800	4*	0	0.7 (1.2)	16.3 (4.2)	17.0 (5.3)	10.3 (5.8)	6.7 (1.2)	35.6	>0.05	43.7	0.018
FQ-OH ^{a,b}	200	3	0	3.7 (5.5)	21.3 (26.6)	25.0 (24.4)	16.3 (14.3)	8.7 (10.8)	17.3	>0.05	33.6	>0.05
RQ ^a	200	4	0	0.25 (0.5)	36.5 (10.7)	36.8 (10.4)	18.8 (5.3)	18.0 (5.3)	0	>0.05	0	>0.05
MQ	200	5	0						72.3 [8]		100	
CQ	200	5	0						11.7 [8]		93.0	

*one mouse died within 24 hours post-treatment, ^aworm burden reduction calculated versus control ¹; ^bworm burden reduction calculated versus control ²SD= standard deviation.

antischistosomal properties of CQ. For comparison, at 200 mg/kg MQ achieved a much higher worm burden reduction of 72.3% in *S. mansoni*-infected mice [8]. A few issues of our *in vivo* results are worth highlighting: interestingly, a moderate female worm burden reduction of 43.7% ($p = 0.018$) was observed using FQ at 800 mg/kg (which had not been observed with CQ). A higher effect against female adult *S. mansoni* was also observed in MQ treated mice [8] pointing to a sex-specific interference of these drugs with the target. Furthermore, in one of the FQ-OH treated mice many dead worms were recovered and a hepatic shift (i.e. worms migrating to the liver) observed. Hence, FQ and FQ-OH show weak antischistosomal activity *in vivo*, which is in line with our *in vitro* results.

Conclusions

In conclusion, the organometallic compounds evaluated in this study show only weak antischistosomal properties *in vivo*. Hence, based on our findings an ancillary benefit on schistosomiasis as a result of treating *P. falciparum* infections with FQ is not expected. Furthermore, no correlation can be drawn between the antimalarial and antischistosomal activity of CQ analogues, which might hint to distinct mechanisms of actions. Despite the low activities of organometallic drugs tested against *S. mansoni* so far [25] further derivatives (e.g. organometallic derivatives of MQ) should be studied.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

JK, GG and CB designed the studies. MV and RR carried out the experiments. JK wrote the first draft of the manuscript. GG and CB revised the manuscript. All authors read and approved the final version of the manuscript.

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Author details

¹Department of Medical Parasitology and Infection Biology, Swiss Tropical and Public Health Institute, Basel, Switzerland. ²University of Basel, Basel, Switzerland. ³Department of Chemistry, University of Zurich, Winterthurerstrasse 190, CH-8057 Zurich, Switzerland. ⁴Université Lille 1, Unité de Glycobiologie Structurale et Fonctionnelle UGSF, F-59650 Villeneuve d'Ascq, France. ⁵CNRS, UMR 8576, F-59650 Villeneuve d'Ascq, France.

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References

1. Colley DG, Bustinduy AL, Secor WE, King CH: Human schistosomiasis. *Lancet* 2014, **383**:2253–2264.

2. World Health Organisation: Schistosomiasis. In *Fact sheet N°115*. Geneva: World Health Organisation; 2014.
3. Utzinger J, N'Goran EK, Caffrey CR, Keiser J: From innovation to application: Social-ecological context, diagnostics, drugs and integrated control of schistosomiasis. *Acta Trop* 2011, **120**(Suppl 1):S121–S137.
4. Keiser J, Utzinger J: Antimalarials in the treatment of schistosomiasis. *Curr Pharm Des* 2012, **18**:3531–3538.
5. Lee H, Moody-Davis A, Saha U, Suzuki BM, Asarnow D, Chen S, Arkin M, Caffrey CR, Singh R: Quantification and clustering of phenotypic screening data using time-series analysis for chemotherapy of schistosomiasis. *BMC Genomics* 2012, **13**(Suppl 1):S4.
6. Abdulla MH, Ruelas DS, Wolff B, Snedecor J, Lim KC, Xu F, Renslo AR, Williams J, McKerrow JH, Caffrey CR: Drug discovery for schistosomiasis: hit and lead compounds identified in a library of known drugs by medium-throughput phenotypic screening. *PLoS Negl Trop Dis* 2009, **3**:e478.
7. Panic G, Duthaler U, Speich B, Keiser J: Repurposing drugs for the treatment and control of helminth infections. *Int J Parasitol Drugs Drug Resist* 2014. doi: 10.1016/j.ijpddr.2014.07.002.
8. Keiser J, Chollet J, Xiao SH, Mei JY, Jiao PY, Utzinger J, Tanner M: Mefloquine-an aminoalcohol with promising antischistosomal properties in mice. *PLoS Negl Trop Dis* 2009, **3**:e350.
9. Keiser J, N'Guessan NA, Adoubryn KD, Silue KD, Vounatsou P, Hatz C, Utzinger J, N'Goran EK: Efficacy and safety of mefloquine, artesunate, mefloquine-artesunate, and praziquantel against *Schistosoma haematobium*: randomized, exploratory open-label trial. *Clin Infect Dis* 2010, **50**:1205–1213.
10. Basra A, Mombongo-Goma G, Melsler MC, Diop DA, Wurbel H, Mackanga JR, Furstenauf M, Zoleko RM, Adegnik AA, Gonzalez R, Menendez C, Kreamsner PG, Ramharther M: Efficacy of mefloquine intermittent preventive treatment in pregnancy against *Schistosoma haematobium* infection in gabon: a nested randomized controlled assessor-blinded clinical trial. *Clin Infect Dis* 2013, **56**:e68–e75.
11. Biot C, Castro W, Botte CY, Navarro M: The therapeutic potential of metal-based antimalarial agents: implications for the mechanism of action. *Dalton Trans* 2012, **41**:6335–6349.
12. Dubar F, Slomianny C, Khalife J, Dive D, Kalamou H, Guerardel Y, Grellier P, Biot C: The ferroquine antimalarial conundrum: redox activation and reinvasion inhibition. *Angew Chem Int Ed Engl* 2013, **52**:7690–7693.
13. Dubar F, Egan TJ, Pradines B, Kuter D, Ncokezi KK, Forge D, Paul JF, Pierrot C, Kalamou H, Khalife J, Buisine E, Rogier C, Veizin H, Forfar I, Slomianny C, Trivelli X, Kaphishnikov S, Leiserowitz L, Dive D, Biot C: The antimalarial ferroquine: role of the metal and intramolecular hydrogen bond in activity and resistance. *ACS Chem Biol* 2011, **6**(3):275–287.
14. Biot C, Daher W, Chavain N, Fandeur T, Khalife J, Dive D, De Clercq E: Design and synthesis of hydroxyferroquine derivatives with antimalarial and antiviral activities. *J Med Chem* 2006, **49**:2845–2849.
15. Keiser J: *In vitro* and *in vivo* trematode models for chemotherapeutic studies. *Parasitology* 2010, **137**:589–603.
16. Biot C, Glorian G, Maciejewski LA, Brocard JS: Synthesis and antimalarial activity *in vitro* and *in vivo* of a new ferrocene-chloroquine analogue. *J Med Chem* 1997, **40**:3715–3718.
17. Manneck T, Haggenmüller Y, Keiser J: Morphological effects and tegumental alterations induced by mefloquine on schistosomula and adult flukes of *Schistosoma mansoni*. *Parasitology* 2010, **137**:85–98.
18. Xiao SH, Keiser J, Chollet J, Utzinger J, Dong Y, Vennerstrom JL, Tanner M: The *in vitro* and *in vivo* activities of synthetic trioxolanes on major human schistosome species. *Antimicrob Agents Chemother* 2007, **51**:1440–1445.
19. Alkadi HO: Antimalarial drug toxicity: a review. *Chemotherapy* 2007, **53**:385–391.
20. Mombongo-Goma G, Supan C, Dal-Bianco MP, Missinou MA, Matsiegui PB, Ospina Salazar CL, Issifou S, Ter-Minassian D, Ramharther M, Kombila M, Kreamsner PG, Lell B: Phase I randomized dose-ascending placebo-controlled trials of ferroquine—a candidate anti-malarial drug—in adults with asymptomatic *Plasmodium falciparum* infection. *Malar J* 2011, **10**:53.
21. Pomel S, Biot C, Bories C, Loiseau PM: Antiprotozoal activity of ferroquine. *Parasitol Res* 2013, **112**:665–669.
22. Biot C, Nosten F, Fraisse L, Ter-Minassian D, Khalife J, Dive D: The antimalarial ferroquine: from bench to clinic. *Parasite* 2011, **18**:207–214.
23. Xue J, Jiang B, Liu CS, Sun J, Xiao SH: Comparative observation on inhibition of hemozoin formation and their *in vitro* and *in vivo* antischistosome activity displayed by 7 antimalarial drugs. *Zhongguo Ji Sheng Chong Xue Yu Ji Sheng Chong Bing Za Zhi* 2013, **31**:161–169.

24. Manneck T, Keiser J, Müller J: Mefloquine interferes with glycolysis in schistosomula of *Schistosoma mansoni* via inhibition of enolase. *Parasitology* 2012, **139**:497–505.
25. Patra M, Ingram K, Leonidova A, Pierroz V, Ferrari S, Robertson MN, Todd MH, Keiser J, Gasser G: *In vitro* metabolic profile and *in vivo* antischistosomal activity studies of (η(6)-praziquantel)Cr(CO)₃ derivatives. *J Med Chem* 2013, **56**:9192–9198.

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